

Bioactive Compounds in Ethanolic Extract of *Strychnos innocua* Root Using Gas Chromatography and Mass Spectrometry (GC-MS)

Alagbe John Olujimi^{1,*}

Abstract

Medicinal plants are of great relevance with endless pharmaceutical and therapeutic properties. They are source of wide array of secondary metabolites or bioactive compounds/phytochemicals (phenols, alkaloids, flavonoids, terpenoids, tannins, steroids and saponins) which can discharge numerous biological functions (antimicrobial, antioxidant, antiviral, antifungal, anti-fibrotic, immune-modulatory, cytotoxic, antipyretic, antitumor, antihelminthic, antiprotozoal, antibacterial, and so on). The use of gas chromatography and mass spectrometry analysis in identifying the bioactive compounds in the ethanolic extract of *Strychnos innocua* root is a fundamental technique in quantifying the unknown samples, trace elements, and contaminants leading to the discovery of novel compounds of pharmaceutical and biomedical importance. The result obtained showed that *Strychnos innocua* root extract contains 39 bioactive compounds with α -cubebene (20.09%) having the highest concentration followed by dibutyl benzene-1,2-dicarboxylate (10.17%), β -elemenone (10.02%), 4-methoxy-2-nitroformanilide (7.21%), 1-methyl cyclopropane methanol (5.96%), 1,3-propanediol, 2-ethyl-2-hydroxymethyl (3.71%), azelaic acid (2.87%), glycidol stearate (2.85%), chloromethyl 2-chlorodecanoate (2.83%) and γ -terpinene (2.56%), respectively. The remaining 29 bioactive compounds have concentrations of less than 2%. It was concluded that all the compounds observed are sources of medication that can be used traditionally in the treatment of human and animal diseases.

Keywords: *Strychnos innocua*, phytochemicals, pharmaceuticals, therapeutics, gas chromatography

INTRODUCTION

Medicinal plants are the major components of all indigenous or alternative systems of medicine. They contain phytochemicals that are safe, non-toxic, and easily affordable [1–3]. According to the World Health Organization (WHO) (1996) [4] about 80% of the population in developing countries rely on medicinal plants for the treatment of various ailments. There are over 2000 medicinal plants with high potential that are yet to be explored [5–7]. Bioactive compounds from these medicinal plants can perform an anti-inflammatory, antifungal, antiviral, antioxidant, immune-stimulatory, analgesic, antibacterial, antiproliferative, cytotoxic, and hepato-protective, antipyretic, antihelminthic, antiprotozoal, anti-depressant, anti-tumor, anti-fibrotic and hypolipidemic properties [8–11] and could also aid in the discovery of drugs [12–15].

Strychnos innocua, also known as Natal orange, belongs to the family Loganiaceae, genus *Strychnos*, and order Gentianales. The tree is found in several countries such as Angola, Guinea,

*Author for Correspondence

Alagbe John Olujimi
E-mail: dralagbe@outlook.com

¹Researcher, Department of Animal Nutrition and Biochemistry, Sumitra Research Institute, Gujarat, India

Received Date: January 08, 2023

Accepted Date: January 17, 2023

Published Date: January 31, 2023

Citation: Alagbe John Olujimi. Bioactive Compounds in Ethanolic Extract of *Strychnos innocua* Root Using Gas Chromatography and Mass Spectrometry (GC-MS). International Journal of Biochemistry and Biomolecule Research. 2023; 1(1): 9–14p.

Madagascar, Malawi, South Africa, Sudan, Mali, Uganda, Malawi, Zimbabwe, Zambia, Ethiopia, and some parts of India [16]. The trees are found in riverine fringes, and sand forests and they can grow up to 3–14 m high with simple leaves characterized by rounded emarginate or subacute apex [17–19]. Extracts from the leaves, roots, and stems can be used traditionally for the treatment of snake bites, gastrointestinal, and skin diseases, pneumonia, and sexually transmitted infections [20–23].

Previous studies have revealed that *Strychnos innocua* leaf, stem, and root extract contains several bioactive compounds with antimicrobial properties and are also capable of inhibiting the activity of some bacteria and fungi such as *Bacillus* spp., *Candida* spp., *Alternaria solani*, *Brevibacillus brevis*, *Cochliobolus lunatus*, *Escherichia coli*, *Enterobacter* spp., *Aspergillus* spp., *Fusarium* spp., *Klebsiella* spp., *Monascus ruber*, *Micrococcus luteus*, *Pseudomonas* spp., *Streptococcus* spp., *Staphylococcus* spp., *Salmonella* spp., and *Shigella shiga*. Phytochemical analysis of methanolic extract from leaves and roots of *Strychnos innocua* revealed the presence of phenolic compounds which are capable of scavenging free radicals (antioxidants) [24–28], preventing the risk of cardiovascular disease [29–32] and performing immune-modulatory activities in animals thus encouraging food safety [33, 34]. This study aimed to evaluate the bioactive compounds in the ethanolic extract of *Strychnos innocua* root using gas chromatography and mass spectrometry (GC-MS).

MATERIALS AND METHODS

Experimental Site, Collection, and Preparation of *Strychnos innocua* Ethanolic Leaf and Root Extract

The study was performed at Sumitra Research Institute, Gujarat, India with a coastline of 1,600 km, 23°13'N 72°41'E. Fresh *Strychnos innocua* root was harvested from Waghai village, Saputara, India, and identified at the Department of Biological Sciences, Sumitra Research Institute, Gujarat, India. The harvested roots were washed with distilled water, dried under the shade for 13 days, blended into powder form with the aid of an electric blender, and kept in an air-tight labeled container. 100 g of grinded sample was soaked in 350 mL of 90% ethanol for 48 hours with occasional stirring thereafter samples were sieved using Whatman's No.1 filter paper (10 cm) and stored in a sterile air-tight container and stored in a cool dry place until further use.

Gas Chromatography and Mass Spectrometry (GC-MS) of Ethanolic Extract from *Strychnos innocua* Root

Gas chromatography-mass spectrometry (GC-MS) analysis of ethanolic extract from *Strychnos innocua* root was performed with a Varian 450 GC system (Model 1842 series, China) equipped with fused silica column and it was operated at a temperature and pressure range of 50°C to 450°C isothermal 1079 PTV injector and 0 to 100 psi, consisting of splitless injector with total flow of 500 ml/minutes at 10 psi, electron range of 150 eV. Secondary compounds were identified as standard compounds in the National Institute of Standards and Technology (NIST).

RESULTS AND DISCUSSION

Medicinal plants contain natural compounds or phytochemicals that are eco-friendly, safe, and locally available with pharmacological properties [35, 36]. They can also be used traditionally for the treatment of various ailments such as colds, coughs, gastrointestinal diseases, skin diseases, respiratory diseases, malaria, typhoid, and snake bites [37, 38]. Bioactive compounds are mostly secondary metabolites produced by plants via subsidiary pathways and are used by plants for growth, or defense against pathogens [39–42]. Secondary metabolites in *Strychnos innocua* ethanolic root extract using gas chromatography and mass spectrometry (GC-MS) (Table 1) reveals that it is largely contains α -cubebene (20.09%), dibutyl benzene-1,2-dicarboxylate (10.17%), β -elemenone (10.02%), 4-methoxy-2-nitroformanilide (7.21%), 1-methyl cyclopropane methanol (5.96%), 1,3 propanediol, 2-ethyl 2-hydroxymethyl (3.71%), azelaic acid (2.87%), glycidol stearate (2.85%), chloromethyl

Table 1. Secondary metabolites in *Strychnos innocua* ethanolic root extract using GC-MS.

Bioactive compounds	Area (%)	R.T. (min)	Functions
Diethyl suberate	1.44	1.450	Antimicrobial and antioxidant
Ethyl oleate	0.72	1.931	Antipyretic and antioxidant
Diisooctyl phthalate	0.01	2.500	Anti-depressant and antifungal
Glycidol stearate	2.85	3.444	Antiviral, hepato-protective and antioxidant
1,2-benzenedicarboxylic acid	1.77		Anti-microbial, anti-proliferative
Monomethyl pimelate	0.02	6.091	Antifungal
γ -terpinene	1.10	9.435	Hepatoprotective and antifungal
4-fluoro-1-methyl-5-carboxylic acid	0.40	10.701	Anti-inflammatory, antibacterial and analgesics
3-Allyl-6-methoxyphenol	1.67	11.331	Antiprotozoal and cytotoxic
Cyclooctane	0.10	15.560	Anti-androgenic, antiviral and anti-inflammatory
Formamide	2.05	15.740	Hepato-protective, hypolipidemic, antimicrobial, and antioxidant
α -cubebene	20.09	15.100	Antibacterial, antifungal, analgesic antipyretic, and antioxidant
2,4,6-octatrien-1-ol	0.77	15.607	Antiviral and antioxidant
9,12-octadecanoic acid	1.06	18.351	Cytotoxic, antioxidant, anti-inflammatory, antitumor, antifungal
α -longipinene	0.15	18.220	Anti-inflammatory, antioxidant, anti-depressant, and antifungal
Azelaic acid	2.87	18.306	Anti-fibrotic, anti-inflammatory and hypolipidemic
Terpinen-4-ol	1.51	18.331	
1,3 propanediol, 2-ethyl 2-hydroxymethyl	3.71	18.211	Antibacterial, anti-inflammatory, antipyretic, antihelminthic and antifungal
γ -terpinene	2.56	19.386	Antioxidant and anti-inflammatory
β -elemenone	10.02	19.931	Cytotoxic and hepato-protective
9-octadecenoic acid	1.16	19.510	Antifungal
Torreyol- α -cadinol	0.83	19.259	Antiviral, hepato-protective and antioxidant
Hepatadec-3-enal	0.30	19.400	Anti-microbial, anti-proliferative, antiviral, antihelminthic and antibacterial
Ethylene diacrylate	0.50	20.209	Analgesics, antibacterial, antifungal
1-hexyl-2-nitrocyclohexane	1.62	21.344	Anti-inflammatory, antioxidant, anti-depressant
Chloromethyl 2-chlorodecanoate	2.83	21.381	Anti-fibrotic, anti-inflammatory
5-methylhexan-2-yl-hepadecyl benzene-1,2 dicarboxylate	1.10	21.100	Antifungal, analgesic antipyretic, and antioxidant
1-methyl cyclopropane methanol	5.96	22.891	Hepatoprotective and antifungal
4-acetoxy-3-methoxystyrene	1.14	23.080	Hepato-protective, hypolipidemic, antimicrobial
4-methoxy-2-nitroformanilide	7.21	23.300	Cytotoxic, antioxidant
α -Terpinolene	0.02	23.701	Antioxidant, anti-proliferative
3-deoxy-d-mannoic acid	1.20	25.186	Antioxidant, antiproliferative, antifungal, and anti-inflammatory
Dibutyl benzene-1,2-dicarboxylate	10.17	25.901	Cytotoxic and hepato-protective
Butyl undecyl benzene-1,2-dicarboxylate	1.31	25.670	Cytotoxic, antioxidant, anti-inflammatory, antitumor, antifungal
9,15-octadecadienoic acid	0.06	26.801	Antibacterial, antifungal, analgesic antipyretic
β -cyclocitral	0.01	28.009	Antitumor, antifungal
α -phellandrene	1.10	34.491	Analgesic antipyretic and antioxidant
β -phenethylamine	0.08	38.567	Anti-bacterial
Humulene	1.12	41.340	Antimicrobial, antifungal, and hypolipidemic
Total	92.59		

R.T.: reaction time (minutes)

2-chlorodecanoate (2.83%) and γ -terpinene (2.56%), respectively. Other compounds reported were less than 2.0% however, they all have marked therapeutic functions (anti-inflammatory, antiviral, antifungal, antioxidant, hypolipidemic, analgesics, antipyretic, cytotoxic, antitumor, and anti-depressant activities) [41, 43]. The GC-MS component analyzed in this study is in consonance with the findings of Hamisu *et al.* (2021) [28] but contrary to the reports of Hoet *et al.* (2006) [44]. These dissimilarities can be ascribed to processing or extraction procedures employed, parts of plant used, species, geographical location, age of plant as well and method of harvesting [44–46]. The presence of phytochemicals in *Strychnos innocua* ethanolic root extract reveals that it can scavenge toxic chemicals in the body, inhibit the activities of pathogenic bacteria in the gut of animals, thus enhancing the absorption of nutrient as well as enhances the activities of enzymes [42, 47, 48].

CONCLUSION

Medicinal plants have several health benefits due to the presence of phytochemicals (alkaloids, flavonoids, phenols, terpenoids, saponins, tannins, and steroids). Analyzing the bioactive components in *Strychnos innocua* ethanolic root extract will unleash some of the potential pharmaceutical properties in the plant. Adopting the use of gas chromatography and mass spectrometry will further aid in identifying and quantifying unknown samples, unknown contaminants, trace elements, and gases.

REFERENCES

1. Olujimi AJ, Muhammad RS, Daniel SM, Christiana OO. Effect of *Trichilia monadelpha* stem bark extract on the fatty acid composition of rabbit's thigh meat. *J Environ Issues Clim Change*. 2022;1(1):63–71.
2. Sharma S, John AO, Xing L, Ram S, Amita K. Comparative analysis of ethanolic *Juniperus thurifera* leaf, stem bark and root extract using gas chromatography and mass spectrometry. *Int J Agric Anim Prod*. 2022;2(6):18–27.
3. Shittu MD, Alagbe JO. Phyto-nutritional profiles of broom weed (*Sida acuta*) leaf extract. *Int J Integr Educ*. 2020;3(11):119–24. doi: 10.31149/ijie.v3i11.845.
4. World Health Organization. TRS 863 - 34th report of the WHO Expert Committee on Specifications for Pharmaceutical Preparations. (WHO Technical Report Series 863). Geneva: World Health Organization; 1996.
5. Alagbe JO, Habiba Z, Adedeji OM, Bamigboye S, Agbonika D. Influence of *Juniperus thurifera* root extract on the nutrient digestibility and caecal microbial count of growing rabbits. *Web of Synergy: International Interdisciplinary Research Journal*. 2022;1(1):5–17.
6. Oluwafemi RA, Agubosi OCP, Alagbe JO. Proximate, minerals, vitamins and amino acid composition of *Prosopis africana* (African mesquite) seed oil. *Asian J Adv Res*. 2021;11(1):21–7.
7. Asuzu CU, Nwosu MO. Studies of the wood of some Nigeria alkaloid-rich *Strychnos* species. *J Hortic*. 2020;12(2):57–62.
8. Agubosi OCP, Alexander J, Alagbe JO. Influence of dietary inclusion of Sunflower (*Helianthus annuus*) oil on growth performance and oxidative status of broiler chicks. *Cent Asian J Med Nat Sci*. 2022;2(7):187–95.
9. Agubosi OCP, Soliu MB, Alagbe JO. Effect of dietary inclusion levels of *Moringa oleifera* oil on the growth performance and nutrient retention of broiler starter chicks. *Cent Asian J Theor Appl Sci*. 2022;3(3):30–9.
10. Olafadehan OA, Oluwafemi RA, Alagbe JO. Performance, haemato-biochemical parameters of broiler chicks administered Rolfe (*Daniellia oliveri*) leaf extract as an antibiotic alternative. *Drug Discov*. 2020;14(33):135–45.
11. Agubosi OCP, Wika BK, Alagbe JO. Effect of dietary inclusion of Sunflower (*Helianthus annuus*) oil on the growth performance of broiler finisher chickens. *Eur J Mod Med Pract*. 2022;2(5):1–10.
12. Shittu MD, Alagbe JO, Ojebiyi OO, Ojediran TK, Rafiu TA. Growth performance and haematological and serum biochemical parameters of broiler chickens given varied concentrations of *Polyalthia longifolia* leaf extract in place of conventional antibiotics. *Anim Sci Genet*. 2022;18(2):57–71.

13. Ruth TN, Anita RL, Loveness KN, Vincenzo F, Ruud V. Local processing and nutritional composition of indigenous fruits: the case of monkey orange (*Strychnos* spp.) from Southern Africa. *Food Res.* 2017;33(2):123–42.
14. Vásquez-Ocmín P, Cojean S, Rengifo E, Suyyagh-Albouz S, Amasifuen Guerra CA, Pomel S et al. Antiprotozoal activity of medicinal plants used by Iquitos-Nauta road communities in Loreto (Peru). *J Ethnopharmacol.* 2018;210:372–85. doi: 10.1016/j.jep.2017.08.039.
15. Hirotani H, Ohigashi H, Kobayashi M, Koshimizu K, Takahashi E. Inactivation of T5 phage by cis-vaccenic acid, an antiviral substance from *Rhodospseudomonas capsulata*, and by unsaturated fatty acids and related alcohols. *FEMS Microbiology Letters.* 1991;77(1):13–8. doi: 10.1111/j.1574-6968.1991.tb04314.x.
16. Prins H, Maghembe JA. Germination studies on seed of fruit trees indigenous to Malawi. *Forest Ecol Manag.* 1994;64(2–3):111–25. doi: 10.1016/0378-1127(94)90285-2.
17. Hines DA, Eckman K. Indigenous multipurpose trees of Tanzania: uses and economic benefits for people. Ottawa, ON, Canada. *Cult Surviv Canada.* 1993.
18. Alagbe JO, Adeoye A, Oluwatobi OA. Proximate and mineral analysis of *Delonix regia* leaves and roots. *Int J Integr Educ.* 2020;3(10):144–9. doi: 10.31149/ijie.v3i10.705.
19. Alagbe JO, Sharma R, Ojo EA, Shittu MD, Atanda BK. Chemical evaluation of the proximate, minerals, vitamins and phytochemical analysis of *Daniellia oliveri* stem bark. *Int J Biol Phys Chem Stud.* 2020;2(1):16–22.
20. Alagbe JO. Caecal microbial population of growing grass cutters (*Thyronoymys swinderianus*) fed *Phyllanthus amarus* and *Pilogstigma Thongii* leaf meal mixture as partial replacement for soya bean meal. *Concept Dairy Vet Sci.* 2020;3(5):350–5.
21. Sun H, Sun C, Pan Y. Cytotoxic Activity and Constituents of the Volatile Oil from the Roots of *Patrinia scabra* Bunge. *C&B.* 2005;2(10):1351–7. doi: 10.1002/cbdv.200590107.
22. Alagbe JO. Gas chromatography and mass spectroscopy of *Juniperus phoenice* stem bark extract and its influence on the haemato-biochemical values of growing rabbits. *Br Sci Period.* 2022;1(1):18–33.
23. Al-Wathnani H, Ismet A, Tahmaz RR, Al-Dayel TH, Bakir MA. Bioactivity of natural compounds isolated from cyanobacteria and green algae against human pathogenic bacteria and yeast. *J Med Plants Res.* 2012;6(18):3425–33. doi: 10.5897/JMPR11.1746.
24. Alagbe JO. *Daniellia oliveri* leaf extracts as an alternative to antibiotic feed additives in broiler chicken diets: meat Quality and fatty acid composition. *Indonesian J Innov Appl Sci.* 2021;1(3):177–86. doi: 10.47540/ijias.v1i3.174.
25. Lee SW, Wendy W, Julius YFS, Desy FS. Characterization of antimicrobial, antioxidant, anticancer properties and chemical composition of *Michelia champaca* seed and flower extracts. *Stamford J Pharm Sci.* 2011;4(1):19–24.
26. Alagbe JO. Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and root of *Indigofera tinctoria*. *Int J Integr Educ.* 2020;3(10):150–7. doi: 10.31149/ijie.v3i10.708.
27. Hussain I. Phytochemicals screening and antimicrobial activities of selected medicinal plants of Khyber Pakhtunkhwa Pakistan. *Afr J Pharm Pharmacol;*5(6):746–50. doi: 10.5897/AJPP11.175.
28. Ibrahim H, Uttu AJ, Sallau MS, Iyun ORA. Gas chromatography–mass spectrometry (GC–MS) analysis of ethyl acetate root bark extract of *Strychnos innocua* (Delile). *Beni Suef Univ J Basic Appl Sci.* 2021;10(1):65. doi: 10.1186/s43088-021-00156-1.
29. Alagbe JO, Shittu MD, Tanimomo BK. Influence of *Anogeissus leiocarpa* stem bark on the fatty acid composition in meat of broiler chickens. *Eur J Life Saf Stab.* 2022;14(22):13–22.
30. Alagbe JO. Use of medicinal plants as a panacea to poultry production and food security: a review. *Am J Technol Appl Sci.* 2022 Apr 26;1:24–36.
31. Alagbe JO. Dietary supplementation of *Rauvolfia vomitoria* root extract as a phytogenic feed additive in growing rabbit diets: growth performance and caecal microbial population. *Concepts Dairy Vet Sci.* 2021;4(2):409–16. doi: 10.32474/CDVS.2021.04.000184.
32. Alagbe JO. *Prosopis africana* stem bark as an alternative to antibiotic feed additives in broiler chicks diets: performance and Carcass characteristics. *J Multidimensional Res Rev.* 2021;2(1):64–77.

-
33. Halliwell B, Gutteridge JM. Free Radicals in Biology and Medicine. Oxford University Press; 2015.
 34. Oloruntola OD, Ayodele SO, Adeyeye SA, Agbede JO. Performance, haemato-biochemical indices and antioxidant status of growing rabbits fed on diets supplemented with *Mucuna pruriens* leaf meal. *World Rabbit Sci.* 2018;26(4):277–85. doi: 10.4995/wrs.2018.10182.
 35. Adewale AO, Alagbe JO, Adeoye AO. Dietary supplementation of *Rauvolfia vomitoria* root extract as a phytogetic feed additive in growing rabbit diets: haematology and serum biochemical indices. *Int J Orange Technol.* 2021;3(3):1–12.
 36. Musa B, Alagbe JO, Adegbite Motunrade B, Omokore EA. Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanites aegyptiaca* and *Alchornea cordifolia* stem bark mixture. *U J Res Technol.* 2020;2(2):13–21.
 37. Nascimento GGF, Locatelli J, Freitas PC, Silva GL. Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Brazil J Microbiol.* 2000;31(4):886–91. doi: 10.1590/S1517–83822000000400003.
 38. Parekh J, Chanda S. In vitro antibacterial activity of the crude methanol extract of *Woodfordia fruticosa* Kurz. Flower (Lythraceae). *Brazil J Microbiol.* 2007;38(2):204–7. doi: 10.1590/S1517–83822007000200004.
 39. Oluwafemi RA, Omolade LA, Adelowo SA, Alagbe JO. Effects of dietary inclusion of ginger (*Zingiber officinale*) and garlic (*Allium sativum*) oil on carcass characteristics and sensory evaluation of broiler chicken. *Texas J Multidiscip Stud.* 2021;2(11):180–8.
 40. Oluwafemi RA, Uankhoba IP, Alagbe JO. Effects of turmeric oil as a dietary supplements on the growth performance and carcass characteristics of broiler chicken. *Int J Orange Technol.* 2021;3(4):1–9.
 41. Okeke MI, Iroegbu CU, Eze EN, Okoli AS, Esimone CO. Evaluation of extracts of the roots of *Landolphia owerience* for antibacterial activity. *J Ethnopharmacology.* 2001;78(2–3):119–27.
 42. Narayani M, Johnson M, Sivaraman A, Janakiraman N. Phytochemical and antibacterial Studies on *Jatropha curcas* L. *J Chem Pharm Res.* 2012;4(5):2639–42.
 43. Trease GE, Evans WC. Pharmacognosy. London: W B Saunders Co Ltd; 1989. p. 269–300.
 44. Hoet S, Stévigny C, Hérent MF, Quetin-Leclercq J. Antitrypanosomal compounds from the leaf essential oil of *Strychnos spinosa*. *Planta Med.* 2006;72(5):480–2. doi: 10.1055/s-2005–916255.
 45. Omokore EO, Alagbe JO. Efficacy of dried *Phyllanthus amarus* leaf meal as an herbal feed additive on the growth performance, haematology and serum biochemistry of growing rabbits. *Int J Acad Res Dev.* 2019;4(3):97–104.
 46. Orwa CA, Mutua KR, Jamnadass R, Anthony S. (2009). Agroforestry database: a tree reference. [online] Available from: <http://www.worldagroforestry.org/sites/treedbs/treedatabases.asp>.
 47. Oluwafemi RA, Uankhoba IP, Alagbe JO. Effects of turmeric oil as a dietary supplement on the haematology and serum biochemical indices of broiler chickens. *Bioinformatics Proteomics Open Access J.* 2021;5(1): 000138.
 48. Oluwafemi RA, Daniel SE, Alagbe JO. Haematology and serum biochemical indices of broiler chicks fed different inclusion levels of ginger (*Zingiber officinale*) and garlic (*Allium sativum*) oil mixture. *Int J Discov Innov Appl Sci.* 2021;1(4):20–6.