

A Narrative Review on Osteogenesis Imperfecta and Its Management

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Abstract

Osteogenesis imperfecta (OI), sometimes referred to as brittle bone disease, is a heterogeneous illness characterized by short stature, several fractures, and distorted bones. The main contributing factor to OI is mutations in the genes needed to produce type 1 collagen. While severe OI is perinatally fatal, mild OI can occasionally not be detected until maturity. The frequency of Osteogenesis imperfecta ranges from about 1:15,000 to 1:20,000 births. Five kinds of the illness are commonly differentiated, ranging in severity from moderate (type I) to deadly (type II). Types III and IV are serious types that permit survival during the newborn period, but type V has a mild to moderate phenotype and interosseous membrane calcification. The majority of the time, mutations in the col I genes result in a decrease in the production of common type I collagen (col I) or the synthesis of uncommon collagen. Moreover, it has been shown that mutations in the genes responsible for osteoblast development, col I processing, and synthesis have occurred. The objective of this study is to review the research on the current medical treatments for osteogenesis imperfecta (OI) in patients. The treatments that are currently available aim to reduce or eliminate symptoms, prevent fractures, and build bone mass. Medications like bisphosphonates, denosumab, and teriparatide are frequently used to treat OI.

Keywords: Osteogenesis imperfecta, bisphosphonates, denosumab, teriparatide, physiotherapy

INTRODUCTION

A collection of inherited skeletal illnesses with a broad range of appearance is called osteogenesis imperfecta (OI), often known as brittle bone disease [1, 2]. The central feature of OI is the skeletal fragility and its predisposition to fractures as a result of structural and or quantitative abnormalities of collagen, especially, type 1 collagen found in bones, teeth tendons, ligaments, skin and sclerae [1]. Several genetic defects have been identified, as well as the different defective proteins. These discoveries have necessitated further genetic classification of OI in addition to an original clinical classification that describes OI syndromes [3–6]. At present, there are five clinical syndromes, which are mostly inherited in an autosomal dominant (AD) pattern and 20 genetic subtypes with various modes of inheritance [5, 7]. The incidence of osteogenesis imperfecta (OI), an uncommon skeletal abnormality,

ranges from 1/15,001 to 1/20,000 [8]. A high incidence of fractures, bone fragility, inadequate growth, and bone abnormalities are kind of Osteogenesis imperfecta [9]. People with OI may also develop other clinical symptoms such brittle teeth, blue sclerae, hearing loss, reduced respiratory function, and heart valve regurgitation because type I collagen formation in numerous tissues is impeded [9]. The most severe form of OI, which is perinatally deadly, can range in intensity from mild to extremely severe [10]. Osteogenesis imperfecta is clinically classified using the Silence scale, which initially identified four categories of Osteogenesis imperfecta based on their clinical presentation [11].

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These “classical” Osteogenesis imperfecta kinds are connected with autosomal mutations in the genes encoding type I collagen (COL1A1, COL1A2) and still account for 85 to 90% of Osteogenesis imperfecta cases [12]. In contrast to structural mutants, haploinsufficiency, which reduces collagen production, often results in milder types of OI. These mutations cause qualitative or structural flaws in the collagen protein [13, 14]. This system of classification has been broadened to incorporate mutations that have been found in a variety of genes that are dominant, recessive, and X-linked. It is possible for non-collagen mutations to cause non-classical OI phenotypes that exhibit distinctive clinical traits, such as the hyperplastic callus formation observed in type V Osteogenesis imperfecta [15]. The present approach to treating Osteogenesis imperfecta concentrates on lowering the risk of fracture, improving motor function, escalating inclusion, and improving quality of life. This requires a multidisciplinary approach from a variety of healthcare providers, including medical doctors, orthopedic surgeons, physiotherapists, geneticists, and other allied health professionals [16]. Antiresorptive medications have been shown in numerous studies to enhance bone mass and lower total risk of fracture, especially in younger populations [17–19]. Notably, medications like bisphosphonates cause increases in bone volume rather than improving the material qualities of bone (i.e., improving bone quality). This results in relative gains in strength.

Classification

OI has historically been categorized in accordance with Sillence, who in the late 1970s established a revised categorization system based on radiological, clinical, and genetic data (Table 1) [20–22]. Sillence type I refers to the mildest and most prevalent kind of OI. Fractures, blue sclerae, and hearing loss are all connected to this kind. These people frequently suffer from many fractures as children, but their clinical conditions improve during puberty and they have a typical life expectancy [23]. The perinatal deadly variety of OI is type II. Infants and fetuses who are affected are frequently stillborn or pass away shortly after birth as a result of many thoracic fractures complicating breathing and perhaps having an intrinsic pulmonary collagen disease [24]. The most serious Osteogenesis imperfecta kind that can endure the newborn period is type III. Numerous fractures may be sustained by OI type III patients, who also frequently have a noticeably small stature, increasing deformities, severe scoliosis, and a condensed lifespan [25–30]. DI is typical, and the hue of the sclera can vary. Sillence type IV is a medium variety, with phenotypes that fall between types I and III in terms of seriousness and clinical traits. The Sillence system of classification is commonly used, although it has currently been complicated by the variation in underlying genetic background and the phenotypic variability of collagen I mutations [31–38]. Every new gene that has been found to create an OI-like phenotype has been given a Roman number, and there are currently 17 types that have been identified (Table 1) [39]. There is no clear agreement on how to identify and categorize this disorder, and the Osteogenesis imperfecta phenotypes brought on by non-collagenous genes overlay with those brought on by conventional dominant OI produced by collagen I mutations [40–49].

PATHOPHYSIOLOGY

The majority of OI patients have an autosomal dominant mutation in COL1A1 or COL1A2, which encode the $\alpha 1(I)$ and $\alpha 2(I)$ chains of type I collagen, respectively. More than 1500 dominant mutations in the COL1A1 and COL1A2 genes have been discovered so far [50–52]. A lack of type I collagen either structurally or quantitatively results from these mutations [52]. Quantitative deficiencies have a milder phenotype than structural and qualitative problems [53]. The most common protein in connective tissue's extracellular matrix is collagen [54]. The triple helix-shaped structure of collagen molecules is made up of three polypeptide chains. For the synthesis of the triple helix, glycine is necessary, as only glycine can fit the partial internal helical space [53]. Type I collagen develops structural and functional flaws when a gene mutation results in glycine replacement, which disrupts helical flossing [55]. In contrast to substitutions in the $\alpha 2(I)$ chains, which are often non-lethal, those in the $\alpha 1(I)$ chains have lethal effects [52]. On the other hand, the mildest form of OI is caused by haploinsufficiency of COL1A1, which reduces the production of structurally normal collagen [56]. A normal phenotype is produced by COL1A2 haploinsufficiency, whereas homozygous null mutations of COL1A2 result in

phenotypes that range from mild to severe Osteogenesis imperfecta. In addition, IFITM5 (interferon inducible transmembrane protein family 5), commonly known as BRIL, has another unusual autosomal dominant mutation [57]. The bone's ability to mineralize and differentiate is inhibited as a result of this mutation [58]. The majority of Osteogenesis imperfecta cases are caused by dominant or sporadic mutations [59]; however, in the past two decades, uncommon autosomal recessive or X-linked mutations have also been discovered [53]. These genes are involved in the osteoblast synthesis (e.g., WNT1, CREB3L1, and SP7), ossification or mineralization (e.g., SERPINF1), extracellular post modification of collagen (e.g., CRTAP, LEPRE1, and PPIB), and collagen folding and intracellular trafficking (e.g., SERPINH1 and FKBP10) [53, 58].

EPIDEMIOLOGY

Osteogenesis imperfecta has a birth rate of 6–7/100,000. The incidence and prevalence of the various OI types vary, with Osteogenesis imperfecta types I and IV making up significantly more than half of all Osteogenesis imperfecta cases [60]. Sillence *et al.* observed a prevalence of 3–4/100,000 and an incidence of 3.5/100,000 for Osteogenesis imperfecta type I in Victoria, Australia. Due to early fatality, data on prevalence for OI type II are not available, however the frequency is approximately 1–2/100,000 [60]. The incidence of OI type III is 1–2/100,000. In Australia, an incidence of 1.6/100,000 has been recorded [3]. OI type IV was thought to be an uncommon phenomenon, but this proved not to be the case [60].

Table 1. Classification of osteogenesis imperfecta types.

Types of Osteogeneses	Phenotype	Gene Affected
Dominant inheritance classical Sillence types		
I	Mild, nondeforming	COL1A1 or COL1A2
II	Perinatal lethal	COL1A1 or COL1A2
III	Progressively deforming	COL1A1 or COL1A2
IV	Moderately deforming	COL1A1 or COL1A2
COL1-mutation negative		
V	Moderate, distinct histology and hyperplastic callus	IFITMS
XV	Mild/early onset osteoporosis	WNT1
Recessive inheritance Mineralization defect		
VI	Moderate to severe, distinct histology	SERPINF1
3-Hydroxylation defects		
VII	Severe to lethal	CRTAP
VIII	Severe to lethal	LEPRE1
IX	Moderate to lethal	PPIB
Chaperone Defect		
X	Severe	SERPINH1
XI	Progressive deforming, Bruck Syndrome	FKBP10
Zinc-finger transcription factor defect		
XII	Moderate	SP7
C-Propeptide cleavage defect		
XIII	Severe high bone mass case	BMP1
Cation Channel defect		
XIV	Moderate to severe	TMEM38B
WNT signaling pathway defect		
XV	Moderate, progressively deforming	WNT1
Unclassified ER stress transducer deficiency X-linked inheritance	Severe	CREB3L1
Unclassified suspected osteocyte defect	Mild	PLS3

MANAGEMENT

Pharmacological Treatment

For OI, current therapy options include managing symptoms, increasing bone mass, and preventing bone fractures. Both non-surgical and surgical techniques are used to treat OI. The non-surgical approach involves the use of medicines, physical therapy, braces, and splints to stay away from deformity and to enhance protection and support. In order to treat local diseases such as scoliosis, bone bending, or fractures, surgery may be necessary. In order to typically combat the systemic effects, medication must be taken. Since the primary publication of the effect of bisphosphonate treatment in children with serious OI, intravenous and oral bisphosphonates are routinely prescribed for all Osteogenesis imperfecta kinds, elders as well as kids [61]. The main justification for bisphosphonate medication is based on several (not placebo-controlled) clinical trials that showed increases in bone mineral density in OI patients. Non-nitrogenous bisphosphonates trigger osteoclast apoptosis, while nitrogenous bisphosphonates interfere with osteoclast development, survival, and cytoskeletal dynamics. According to a newly published systematic evaluation of bisphosphonate treatment in Osteogenesis imperfecta [62], there is a considerable improvement in bone mineral density in a relatively limited proportion of patients who have OI and are receiving intravenous or oral bisphosphonates. The most necessary query that has arisen, however, is whether rise in bone mineral density leads to a decrease in fractures and an improvement in function; this subject has not yet been addressed and calls for further research [62]. The use of growth hormone to reduce short stature in types III and IV Osteogenesis imperfecta [63] is presently being researched [64]. Denosumab is a monoclonal antibody (IgG2) that targets RANKL, a nuclear factor kappa-B ligand that inhibits the synthesis of osteoclasts without attaching to bone [65]. The benefit of Denosumab is a relatively brief degradation period, which lasts for 3 to 4 months, avoiding the long-term accumulating negative effects of BPs [66]. This compound has demonstrated promising results with a comparatively high level of safety when investigated in patients with OI types I, III, IV, and VI who are not sensitive to BPs [66, 67]. Additionally, after 10 years of therapy with Denosumab in postmenopausal women with osteoporosis, studies from the major, phase III clinical FREEDOM trial showed persistent increases in bone mineral density with only low rates of side events and a low fracture incidence [67, 68]. In addition, there have been reports of hypercalcemia, hypercalciuria, and rebound symptoms after quitting the medication for Denosumab [69]. These reports suggest that more research on this substance is still needed. In a randomized controlled trial, the bone anabolic drug teriparatide was evaluated in people with Osteogenesis imperfecta [70]. In mild Osteogenesis imperfecta, there was an increase in BMD, but it was less effective in moderate-to-severe Osteogenesis imperfecta. According to these results, teriparatide showed beneficial effects in two studies that targeted individuals with Osteogenesis imperfecta type I [71, 72]. Teriparatide may therefore be effective in the management of mild Osteogenesis imperfecta in elders. However, teriparatide should not be used in youngsters, though, as research in developing rats has linked it to an elevated risk of osteogenic sarcoma [73].

Gene and Cell Therapy

The two ways by which mutations are reduced in nature, silencing a mutant allele and cellular mosaicism, are emulated in the most potential future treatments for OI caused by faulty collagen structure. To mimic the scenario in type I OI where the mutant allele is quiet, many gene-silencing approaches (ribozymes, small interfering RNA, and short hairpin RNA) have been examined *in vitro* [74]. Instead of curing OI, this strategy would cause a milder OI. Although these methods can produce silencing that is nearly complete in a test tube, the silencing in cells is typically incomplete, and the duration of the impact is constrained. The condition in mosaic parents of Osteogenesis imperfecta kids, who have two populations of cells (nonmutant and mutant), and rarely seek medical assistance before the birth of their child, is mimicked by cell transplantation. Cell absorption into bone has been a problem for this strategy. Despite stem cell uptake being less than a little percentage points, studies in mouse models used enhanced bone mechanics [75, 76]. Green fluorescence protein (GFP)-tagged cells have been retransplanted to a secondary recipient, demonstrating their survival, and higher levels of cell transplantation have been achieved with local transplantation more recently [77]. Fetal mesenchymal

stem cells are being transplanted *in utero* or during the first few months after birth in a clinical experiment now taking place in Europe.

Surgical treatment

Reducing pain and ensuring recovery without a rise in abnormalities are the two main goals of orthopedic operations in the treatment of patients with OI. Conservative therapy with a cast is frequently sufficient for fractures without dislocation or in individuals whose bone is aided by an intramedullar rod. Children who have severe long bone abnormalities that might prevent verticalization or limit the functional use of the higher extremity should undergo surgical therapy to repair the deformities. Intramedullar telescopic rods ought to be implanted throughout growing. These grow longer during development and can support the bones for a very long time [78]. It should be highlighted that extended immobilization is not necessary for individuals with classical Osteogenesis imperfecta caused by mutations in COL1A1/2 because these patients' fractures heal just as quickly as those in unaffected people. Healing might be modified in some uncommon types. For instance, this may occur in individuals with mutations in IFITM5 (hyperplastic callus formation) or WNT1 (delayed healing). To avoid muscular atrophy and subsequent bone resorption brought on by immobility, prolonged immobilization must be avoided.

Physiotherapy Management

From the time an OI individual is diagnosed, the physiotherapist plays a critical part in their care. Depending on the seriousness of each individual condition, they can help parents of early Osteogenesis imperfecta babies and kids with safe positioning and handling of the child and setting realistic aims for achieving developmental milestones. Among these milestones, the capacity of the child to move independently, whether with help from a walker or later a wheelchair, is probably the most important. To improve the musculoskeletal health of children with Osteogenesis imperfecta and support their transition into independent adulthood, the physiotherapist's long-term engagement is essential. There is proof from observational research that supervised exercise programs can improve physical capacity and muscle strength [79, 80]. According to anecdotal evidence, strengthening muscles may help individual with hypermobility, a typical symptom of Osteogenesis imperfecta, stabilize their joints. Persons with Osteogenesis imperfecta have a propensity to be discouraged from engaging in sports, yet there are several instances of persons with Osteogenesis imperfecta succeeding in their chosen sports. This can then enhance quality of life and general well-being. Many individuals are hesitant to try novel movement patterns due to recurrent fractures, which should be taken into account throughout training. Additionally, as muscles get stronger, an osteoanabolic signal is triggered, which causes osteoblasts to produce more extracellular matrix. Utilizing the muscles is still the greatest strategy to induce bone growth even though OI can decrease osteoblast activity [81].

CONCLUSION

OI has evolved over the past few decades from a condition with an unidentified etiology to a disorder with a thoroughly characterized and mapped genetic background. Medications such as bisphosphonates, denosumab, and teriparatide are used to treat people with OI; among these, bisphosphonates are the most promising therapy for promoting bone formation by inhibiting bone resorption. Surgery guarantees a reduction in pain and recovery without worsening of abnormalities. Timely remobilization and consistent muscle strengthening through physiotherapy are crucial for improving mobility, preventing bone resorption, and avoiding muscle wasting due to immobilization. However, the majority of these strategies are still in the testing phase. Further research is therefore required to verify their therapeutic advantages in osteogenesis imperfecta.

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Declaration of Competing Interest

None.

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Data sharing does not apply to this article as no new data were created or analyzed in this study.

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REFERENCES

1. Gupta S. Differential Diagnosis in Pediatrics: Including Color Atlas. 5th edition. New Delhi: Jaypee Bros. Medical Publishers; 2009.
2. Mortier GR, Cohn DH, Cormier-Daire V, Hall C, Krakow D, Mundlos S, Nishimura G, Robertson S, Sangiorgi L, Savarirayan R, Sillence D. Nosology and classification of genetic skeletal disorders: 2019 revision. *Am J Med Genet A*. 2019 Dec; 179(12): 2393–419.
3. Sillence D, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. *J Med Genet*. 1979 Apr 1; 16(2): 101–16.
4. Van Dijk FS, Sillence DO. Osteogenesis imperfecta: clinical diagnosis, nomenclature and severity assessment. *Am J Med Genet A*. 2014 Jun; 164(6): 1470–81.
5. Chetty M, Roomaney IA, Beighton P. The evolution of the nosology of osteogenesis imperfecta. *Clin Genet*. 2021 Jan; 99(1): 42–52.
6. Ango S, Fatunke ET, Jemide OO. Osteogenesis Imperfecta in a Newborn: Case Report and Review of the Literature. *Nigerian Journal of Perinatal and Neonatal Care (NJPNC)*. 2023 Mar 14; 3(1): 40–6.
7. Marom R, Rabenhorst BM, Morello R. Management of endocrine disease: Osteogenesis imperfecta: An update on clinical features and therapies. *Eur J Endocrinol*. 2020 Oct 1; 183(4): R95–106.
8. Bregou Bourgeois A, Aubry-Rozier B, Bonafé L, Laurent-Applegate LA, Pioletti D, Zambelli PY. Osteogenesis imperfecta: from diagnosis and multidisciplinary treatment to future perspectives. *Swiss Med Wkly*. 2016; 146: w14322.
9. Forlino A, Marini JC. Osteogenesis imperfecta. *Lancet*. 2016 Apr 16; 387(10028): 1657–71.
10. Götherström C, Westgren M, Shaw SS, Åström E, Biswas A, Byers PH, Mattar CN, Graham GE, Taslimi J, Ewald U, Fisk NM. Pre- and postnatal transplantation of fetal mesenchymal stem cells in osteogenesis imperfecta: a two-center experience. *Stem Cells Transl Med*. 2014 Feb 1; 3(2): 255–64.
11. Sillence D, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. *J Med Genet*. 1979 Apr 1; 16(2): 101–16.
12. Bardai G, Moffatt P, Glorieux FH, Rauch F. DNA sequence analysis in 598 individuals with a clinical diagnosis of osteogenesis imperfecta: diagnostic yield and mutation spectrum. *Osteopor Int*. 2016 Dec; 27(12): 3607–13.
13. Robinson ME, Rauch F. Mendelian bone fragility disorders. *Bone*. 2019 Sep 1; 126: 11–7.

14. Forlino A, Marini JC. Osteogenesis imperfecta. *Lancet*. 2016 Apr 16;387(10028):1657-71. doi: 10.1016/S0140-6736(15)00728-X.
15. Cheung MS, Glorieux FH, Rauch F. Natural history of hyperplastic callus formation in osteogenesis imperfecta type V. *J Bone Miner Res*. 2007 Aug; 22(8): 1181–6.
16. Marr C, Seasman A, Bishop N. Managing the patient with osteogenesis imperfecta: a multidisciplinary approach. *J Multidiscip Healthc*. 2017 Apr 4; 10: 145–55.
17. Bains JS, Carter EM, Citron KP, Boskey AL, Shapiro JR, Steiner RD, Smith PA, Bober MB, Hart T, Cuthbertson D, Krischer J. A multicenter observational cohort study to evaluate the effects of bisphosphonate exposure on bone mineral density and other health outcomes in osteogenesis imperfecta. *JBM Plus*. 2019 May; 3(5): e10118.
18. Lv F, Liu Y, Xu X, Song Y, Li L, Jiang Y, Wang O, Xia W, Xing X, Li M. Zoledronic acid versus alendronate in the treatment of children with osteogenesis imperfecta: a 2-year clinical study. *Endocr Pract*. 2018 Feb 1; 24(2): 179–88.
19. Li LJ, Zheng WB, Zhao DC, Yu W, Wang O, Jiang Y, Xia WB, Li M. Effects of zoledronic acid on vertebral shape of children and adolescents with osteogenesis imperfecta. *Bone*. 2019 Oct 1; 127: 164–71.
20. Sillence DO, Rimo DL, Danks DM. Clinical variability in osteogenesis imperfecta-variable expressivity or genetic heterogeneity. *Birth Defects Orig Artic Ser*. 1979 Jan 1; 15(5B): 113–29.
21. Sillence D, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. *J Med Genet*. 1979 Apr 1; 16(2): 101–16.
22. Sillence D, Rimo DL. Classification of osteogenesis imperfecta. *Lancet*. 1978 May 13; 311(8072): 1041–2.
23. McAllion SJ, Paterson CR. Causes of death in osteogenesis imperfecta. *J Clin Pathol*. 1996 Aug 1; 49(8): 627–30.
24. Thibeault DW, Pettett G, Mabry SM, Rezaiekhaliq MM. Osteogenesis imperfecta Type IIA and pulmonary hypoplasia with normal alveolar development. *Pediatr Pulmonol*. 1995 Nov; 20(5): 301–6.
25. Sykes B, Wordsworth P, Ogilvie D, Anderson J, Jones N. Osteogenesis imperfecta is linked to both type I collagen structural genes. *Lancet*. 1986 Jul 12; 328(8498): 69–72.
26. Morello R, Bertin TK, Chen Y, Hicks J, Tonachini L, Monticone M, Castagnola P, Rauch F, Glorieux FH, Vranka J, Bächinger HP. CRTAP is required for prolyl 3-hydroxylation and mutations cause recessive osteogenesis imperfecta. *Cell*. 2006 Oct 20; 127(2): 291–304.
27. Barnes AM, Chang W, Morello R, Cabral WA, Weis M, Eyre DR, Leikin S, Makareeva E, Kuznetsova N, Uveges TE, Ashok A. Deficiency of cartilage-associated protein in recessive lethal osteogenesis imperfecta. *New Engl J Med*. 2006 Dec 28; 355(26): 2757–64.
28. Cabral WA, Chang W, Barnes AM, Weis M, Scott MA, Leikin S, Makareeva E, Kuznetsova NV, Rosenbaum KN, Tifft CJ, Bulas DI. Prolyl 3-hydroxylase 1 deficiency causes a recessive metabolic bone disorder resembling lethal/severe osteogenesis imperfecta. *Nat Genet*. 2007 Mar; 39(3): 359–65.
29. Barnes AM, Carter EM, Cabral WA, Weis M, Chang W, Makareeva E, Leikin S, Rotimi CN, Eyre DR, Raggio CL, Marini JC. Lack of cyclophilin B in osteogenesis imperfecta with normal collagen folding. *New Engl J Med*. 2010 Feb 11; 362(6): 521–8.
30. van Dijk FS, Nesbitt IM, Zwikstra EH, Nikkels PG, Piersma SR, Fratantoni SA, Jimenez CR, Huizer M, Morsman AC, Cobben JM, van Roij MH. PPIB mutations cause severe osteogenesis imperfecta. *Am J Hum Genet*. 2009 Oct 9; 85(4): 521–7.
31. Baldridge D, Schwarze U, Morello R, Lenington J, Bertin TK, Pace JM, Pepin MG, Weis M, Eyre DR, Walsh J, Lambert D. CRTAP and LEPRE1 mutations in recessive osteogenesis imperfecta. *Hum Mutat*. 2008 Dec; 29(12): 1435–42.
32. Alanay Y, Avaygan H, Camacho N, Utine GE, Boduroglu K, Aktas D, Alikasifoglu M, Tuncbilek E, Orhan D, Bakar FT, Zabel B. Mutations in the gene encoding the RER protein FKBP65 cause autosomal-recessive osteogenesis imperfecta. *Am J Hum Genet*. 2010 Apr 9; 86(4): 551–9.

33. Kelley BP, Malfait F, Bonafe L, Baldridge D, Homan E, Symoens S, Willaert A, Elcioglu N, Van Maldergem L, Verellen-Dumoulin C, Gillerot Y. Mutations in FKBP10 cause recessive osteogenesis imperfecta and Bruck syndrome. *J Bone Miner Res.* 2011 Mar; 26(3): 666–72.
34. Christiansen HE, Schwarze U, Pyott SM, AlSwaid A, Al Balwi M, Alrasheed S, Pepin MG, Weis MA, Eyre DR, Byers PH. Homozygosity for a missense mutation in SERPINH1, which encodes the collagen chaperone protein HSP47, results in severe recessive osteogenesis imperfecta. *Am J Hum Genet.* 2010 Mar 12; 86(3): 389–98.
35. Becker J, Semler O, Gilissen C, Li Y, Bolz HJ, Giunta C, Bergmann C, Rohrbach M, Koerber F, Zimmermann K, de Vries P. Exome sequencing identifies truncating mutations in human SERPINF1 in autosomal-recessive osteogenesis imperfecta. *Am J Hum Genet.* 2011 Mar 11; 88(3): 362–71.
36. Homan EP, Rauch F, Grafe I, Lietman C, Doll JA, Dawson B, Bertin T, Napierala D, Morello R, Gibbs R, White L. Mutations in SERPINF1 cause osteogenesis imperfecta type VI. *J Bone Miner Res.* 2011 Dec; 26(12): 2798–803.
37. Lapunzina P, Aglan M, Temtamy S, Caparrós-Martín JA, Valencia M, Letón R, Martínez-Glez V, Elhossini R, Amr K, Vilaboa N, Ruiz-Perez VL. Identification of a frameshift mutation in Osterix in a patient with recessive osteogenesis imperfecta. *Am J Hum Genet.* 2010 Jul 9; 87(1): 110–4.
38. Martínez-Glez V, Valencia M, Caparrós-Martín JA, Aglan M, Temtamy S, Tenorio J, Pulido V, Lindert U, Rohrbach M, Eyre D, Giunta C. Identification of a mutation causing deficient BMP1/mTLD proteolytic activity in autosomal recessive osteogenesis imperfecta. *Hum Mutat.* 2012 Feb; 33(2): 343–50.
39. Laine CM, Joeng KS, Campeau PM, Kiviranta R, Tarkkonen K, Grover M, Lu JT, Pekkinen M, Wessman M, Heino TJ, Nieminen-Pihala V. WNT1 mutations in early-onset osteoporosis and osteogenesis imperfecta. *New Engl J Med.* 2013 May 9; 368(19): 1809–16.
40. Cho TJ, Lee KE, Lee SK, Song SJ, Kim KJ, Jeon D, Lee G, Kim HN, Lee HR, Eom HH, Lee ZH. A single recurrent mutation in the 5'-UTR of IFITM5 causes osteogenesis imperfecta type V. *Am J Hum Genet.* 2012 Aug 10; 91(2): 343–8.
41. Shapiro JR, Lietman C, Grover M, Lu JT, Nagamani SC, Dawson BC, Baldridge DM, Bainbridge MN, Cohn DH, Blazo M, Roberts TT. Phenotypic variability of osteogenesis imperfecta type V caused by an IFITM5 mutation. *J Bone Miner Res.* 2013 Jul; 28(7): 1523–30.
42. Shaheen R, Alazami AM, Alshammari MJ, Fageih E, Alhashmi N, Mousa N, Alsinani A, Ansari S, Alzahrani F, Al-Owain M, Alzayed ZS. Study of autosomal recessive osteogenesis imperfecta in Arabia reveals a novel locus defined by TMEM38B mutation. *J Med Genet.* 2012 Oct 1; 49(10): 630–5.
43. Volodarsky M, Markus B, Cohen I, Staretz-Chacham O, Flusser H, Landau D, Shelef I, Langer Y, Birk OS. A Deletion Mutation in TMEM 38 B Associated with Autosomal Recessive Osteogenesis Imperfecta. *Hum Mutat.* 2013 Apr; 34(4): 582–6.
44. van Dijk FS, Zillikens MC, Micha D, Riessland M, Marcelis CL, de Die-Smulders CE, Milbradt J, Franken AA, Harsevoort AJ, Lichtenbelt KD, Pruijs HE. PLS3 mutations in X-linked osteoporosis with fractures. *New Engl J Med.* 2013 Oct 17; 369(16): 1529–36.
45. Symoens S, Malfait F, D'hondt S, Callewaert B, Dheedene A, Steyaert W, Bächinger HP, De Paepe A, Kayserili H, Coucke PJ. Deficiency for the ER-stress transducer OASIS causes severe recessive osteogenesis imperfecta in humans. *Orphanet J Rare Dis.* 2013 Dec; 8: 1–6.
46. Pyott SM, Tran TT, Leistriz DF, Pepin MG, Mendelsohn NJ, Temme RT, Fernandez BA, Elsayed SM, Elsobky E, Verma I, Nair S. WNT1 mutations in families affected by moderately severe and progressive recessive osteogenesis imperfecta. *Am J Hum Genet.* 2013 Apr 4; 92(4): 590–7.
47. Pyott SM, Schwarze U, Christiansen HE, Pepin MG, Leistriz DF, Dineen R, Harris C, Burton BK, Angle B, Kim K, Sussman MD. Mutations in PPIB (cyclophilin B) delay type I procollagen chain association and result in perinatal lethal to moderate osteogenesis imperfecta phenotypes. *Hum Mol Genet.* 2011 Apr 15; 20(8): 1595–609.
48. Marini JC, Cabral WA, Barnes AM. Null mutations in LEPRE1 and CRTAP cause severe recessive osteogenesis imperfecta. *Cell Tissue Res.* 2010 Jan; 339(1): 59–70.

49. Marini JC, Blissett AR. New genes in bone development: what's new in osteogenesis imperfecta. *J Clin Endocrinol Metab.* 2013 Aug 1; 98(8): 3095–103.
50. Byers PH, Pyott SM. Recessively inherited forms of osteogenesis imperfecta. *Annu Rev Genet.* 2012 Dec 15; 46: 475–97.
51. Forlino A, Cabral WA, Barnes A, Marini C. New perspectives on osteogenesis imperfecta. *Nat Rev Endocrinol.* 2011 Jun 14; 7(9): 540–557.
52. Marini JC, Forlino A, Cabral WA, Barnes AM, San Antonio JD, Milgrom S, Hyland JC, Körkkö J, Prockop DJ, De Paepe A, Coucke P. Consortium for osteogenesis imperfecta mutations in the helical domain of type I collagen: regions rich in lethal mutations align with collagen binding sites for integrins and proteoglycans. *Hum Mutat.* 2007 Mar; 28(3): 209–21.
53. Forlino A, Marini JC. Osteogenesis imperfecta. *Lancet.* 2016 Apr 16; 387(10028): 1657–71.
54. Viguet-Carrin S, Garnero P, Delmas PD. The role of collagen in bone strength. *Osteoporos Int.* 2006 Mar; 17(3): 319–36.
55. Rauch F, Lalic L, Roughley P, Glorieux FH. Relationship between genotype and skeletal phenotype in children and adolescents with osteogenesis imperfecta. *J Bone Miner Res.* 2010 Jun; 25(6): 1367–74.
56. Willing MC, Deschenes SP, Slayton RL, Roberts EJ. Premature chain termination is a unifying mechanism for COL1A1 null alleles in osteogenesis imperfecta type I cell strains. *Am J Hum Genet.* 1996 Oct; 59(4): 799–809.
57. Hoyer-Kuhn H, Semler O, Schoenau E. Effect of denosumab on the growing skeleton in osteogenesis imperfecta. *J Clin Endocrinol Metab.* 2014 Nov 1; 99(11): 3954–5.
58. Marom R, Lee YC, Grafe I, Lee B. Pharmacological and biological therapeutic strategies for osteogenesis imperfecta. In *American Journal of Medical Genetics Part C: Seminars in Medical Genetics.* 2016 Dec; 172(4): 367–383.
59. Krakow D. Skeletal dysplasias. *Clin Perinatol.* 2015 Jun 1; 42(2): 301–19.
60. Pagon RA, Bird TD, Dolan CR, Stephens K, Waters AM, Beales PL. Bardet–Biedl syndrome. *GeneReviews* (®). Seattle, WA: University of Washington, Seattle; 1993.
61. Glorieux FH, Bishop NJ, Plotkin H, Chabot G, Lanoue G, Travers R. Cyclic administration of pamidronate in children with severe osteogenesis imperfecta. *New Engl J Med.* 1998 Oct 1; 339(14): 947–52.
62. Dwan K, Phillipi CA, Steiner RD, Basel D. Bisphosphonate therapy for osteogenesis imperfecta. *Cochrane Database Syst Rev.* 2016;10:CD005088. DOI: 10.1002/14651858.CD005088.pub4.
63. Marini JC, Hopkins E, Glorieux FH, Chrousos GP, Reynolds JC, Gundberg CM, Reing CM. Positive linear growth and bone responses to growth hormone treatment in children with types III and IV osteogenesis imperfecta: high predictive value of the carboxyterminal propeptide of type I procollagen. *J Bone Miner Res.* 2003 Feb; 18(2): 237–43.
64. van Dijk FS, Cobben JM, Kariminejad A, Maugeri A, Nikkels PG, van Rijn RR, Pals G. Osteogenesis imperfecta: a review with clinical examples. *Mol Syndromol.* 2011 Dec 1; 2(1): 1–20.
65. Narayanan P. Denosumab: A comprehensive review. *South Asian J Cancer.* 2013 Oct; 2(04): 272–7.
66. Li G, Jin Y, Levine MA, Hoyer-Kuhn H, Ward L, Adachi JD. Systematic review of the effect of denosumab on children with osteogenesis imperfecta showed inconsistent findings. *Acta Paediatr.* 2018 Mar; 107(3): 534–7.
67. Hoyer-Kuhn H, Franklin J, Allo G, Kron M, Netzer C, Eysel P, Hero B, Schoenau E, Semler O. Safety and efficacy of denosumab in children with osteogenesis imperfecta-a first prospective trial. *J Musculoskelet Neuronal Interact.* 2016 Mar; 16(1): 24–32.
68. Bone HG, Wagman RB, Brandi ML, Brown JP, Chapurlat R, Cummings SR, Czerwiński E, Fahrleitner-Pammer A, Kendler DL, Lippuner K, Reginster JY. 10 years of denosumab treatment in postmenopausal women with osteoporosis: results from the phase 3 randomised FREEDOM trial and open-label extension. *Lancet Diabetes Endocrinol.* 2017 Jul 1; 5(7): 513–23.
69. Morello R. Osteogenesis imperfecta and therapeutics. *Matrix Biol.* 2018 Oct 1; 71: 294–312.

70. Orwoll ES, Shapiro J, Veith S, Wang Y, Lapidus J, Vanek C, Reeder JL, Keaveny TM, Lee DC, Mullins MA, Nagamani SC. Evaluation of teriparatide treatment in adults with osteogenesis imperfecta. *J Clin Invest*. 2014 Feb 3; 124(2): 491–8.
71. Leali PT, Balsano M, Maestretti G, Brusoni M, Amorese V, Ciurlia E, Andreozzi M, Caggiari G, Doria C. Efficacy of teriparatide vs neridronate in adults with osteogenesis imperfecta type I: a prospective randomized international clinical study. *Clin Cases Miner Bone Metab*. 2017 May; 14(2): 153–156.
72. Gatti D, Rossini M, Viapiana O, Povino MR, Liuzza S, Fracassi E, Idolazzi L, Adami S. Teriparatide treatment in adult patients with osteogenesis imperfecta type I. *Calcif Tissue Int*. 2013 Nov; 93(5): 448–52.
73. Vahle JL, Sato M, Long GG, Young JK, Francis PC, Engelhardt JA, Westmore MS, Ma YL, Nold JB. Skeletal changes in rats given daily subcutaneous injections of recombinant human parathyroid hormone (1-34) for 2 years and relevance to human safety. *Toxicol Pathol*. 2002 Apr; 30(3): 312–21.
74. Besio R, Forlino A. New frontiers for dominant osteogenesis imperfecta treatment: gene/cellular therapy approaches. *Advances in Regenerative Biology (ARB)*. 2015 Jan 1; 2(1): 27964.
75. Guillot PV, Abass O, Bassett JD, Shefelbine SJ, Bou-Gharios G, Chan J, Kurata H, Williams GR, Polak J, Fisk NM. Intrauterine transplantation of human fetal mesenchymal stem cells from first-trimester blood repairs bone and reduces fractures in osteogenesis imperfecta mice. *Blood, The Journal of the American Society of Hematology*. 2008 Feb 1; 111(3): 1717–25.
76. Panaroni C, Gioia R, Lupi A, Besio R, Goldstein SA, Kreider J, Leikin S, Vera JC, Mertz EL, Perilli E, Baruffaldi F. In utero transplantation of adult bone marrow decreases perinatal lethality and rescues the bone phenotype in the knock in murine model for classical, dominant osteogenesis imperfecta. *Blood, The Journal of the American Society of Hematology*. 2009 Jul 9; 114(2): 459–68.
77. Sinder BP, Novak S, Wee NK, Basile M, Maye P, Matthews BG, Kalajzic I. Engraftment of skeletal progenitor cells by bone-directed transplantation improves osteogenesis imperfecta murine bone phenotype. *Stem Cells*. 2020 Apr 1; 38(4): 530–41.
78. Wirth T. The orthopaedic management of long bone deformities in genetically and acquired generalized bone weakening conditions. *J Child Orthop*. 2019 Feb; 13(1): 12–21.
79. Binder H, Conway A, Gerber LH. Rehabilitation approaches to children with osteogenesis imperfecta: a ten-year experience. *Arch Phys Med Rehabil*. 1993 Apr 1; 74(4): 386–90.
80. Binder H, Conway A, Hason S, Gerber LH, Marini J, Berry R, Weintrob J. Comprehensive rehabilitation of the child with osteogenesis imperfecta. *Am J Med Genet*. 1993 Jan 15; 45(2): 265–9.
81. Mueller B, Engelbert R, Baratta-Ziska F, Bartels B, Blanc N, Brizola E, Fraschini P, Hill C, Marr C, Mills L, Montpetit K. Consensus statement on physical rehabilitation in children and adolescents with osteogenesis imperfecta. *Orphanet J Rare Dis*. 2018 Dec; 13(1): 158(4p).