

# Molecular Docking of *Nigella Sativa* Phytochemical as Inhibitors of Transcription Factors NF-KB Implicated in Rheumatoid Arthritis

Amaani\*

## Abstract

**Objectives:** *Nigella sativa*, a plant considered around the world as the most treasured nutrient-rich herb for centuries together in different civilizations, is exceptionally known for its high levels of antioxidant properties. Free radical intensification due to oxidative stress plays a key role in the pathogenesis of rheumatoid arthritis by activating the NF-KB protein that regulates the expression of the genes involved in inflammation. **Methods:** This study involves in-silico approach to validate the antioxidant properties of the phytochemicals obtained from *Nigella sativa* in inhibiting the protein activity of NF-KB by contemplating the binding affinity of the phytochemicals with the target macromolecule virtually by Molecular Docking (PyRx). **Results:** Phytochemical compounds that surpassed the ADME test were screened based on the binding affinity against the protein. Ligands showing values less than -6 kcal/mol were further visualized, and the computation results manifested phytochemical Nigellone (Dithymoquinone) had the strongest binding affinity against the protein NF-KB, recommending the compound for future therapeutic interventions in the treatment of RA.

**Keywords:** *Nigella sativa*, Nigellone, NF-KB, free radicals, oxidative stress, rheumatoid arthritis

## INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory autoimmune disease (AID) of the synovial membrane causing progressive and irreversible joint destruction left untreated, which leads to ankylosis [1]. RA can affect an individual at any age, predominantly occurring 2–3 times more in women. In 2020, an estimated 17.6 million people were affected globally, taking over 0.5–1% of the population, and it is forecasted to be taking over 31.7 million people worldwide by 2050 [2]. The Incident rates in the Indian populations are estimated to be 0.9% [3], in North Africa and the Middle East around 0.16% [3, 4], whereas in US reportedly 0.5–1% [4, 5]. RA is a progressive disease typically occurring in stages over a span of multiple years. The HLA-DRB1 genetic risk locus, the shared epitope (SE) alleles (conserved sequences of 5 amino acids), is the key genetic risk factor strongly associated with the development of RA [6, 7, 8]. In the initial stages of RA, malfunctioning mitochondria and increased DNA damage sustain the free radical generation in the synovium, causing oxidative stress (OS), which

has shown a direct correlation associated with synovitis causing degeneration of the cartilage by denaturation of collagen and proteoglycan protein [1, 9].

Reactive oxygen species (ROS) in the immune system act as a signaling molecule and are observed to be at alleviated levels in patients with RA [10]. RA Pathogenesis is associated with decreased antioxidant levels, impaired antioxidant defenses, and increased oxidative stress, which leads to major consequences like lipid peroxidation. In spite of the cellular repair systems ability to correct most of

### \*Author for Correspondence

Amaani

E-mail: [amaanishk@gmail.com](mailto:amaanishk@gmail.com)

Student, Department of Bioinformatics, BioNome, Hennur Gardens, Bengaluru, India

Received Date: August 30, 2024

Accepted Date: October 28, 2024

Published Date: November 18, 2024

**Citation:** Amaani. Molecular Docking of *Nigella Sativa* Phytochemical as Inhibitors of Transcription Factors NF-KB Implicated in Rheumatoid Arthritis. International Journal of Molecular Biotechnological and Research. 2024; 2(2): 42–55p.

these damages, DNA lesions caused due to free radicals over a prolonged period could be an important attribute towards understanding autoimmune diseases like RA [1]. One such transcription factor (TF) protein activated by free radicals (ROS) is nuclear factor-kappa B (NF-KB) P52 bound to DNA, which plays a major role in the pathogenesis of RA by regulating the expressions of genes involved in immune response, cell survival, and inflammation [11–13].

The main objective of this study is to inhibit the mechanism of free radicals and downregulate the functioning of NF-KB in the cellular destruction of RA by effectively utilizing the antioxidant properties of the bioactive compound obtained from *Nigella sativa* nigellone over NF-KB [14–16]. Current treatment plans include non-specific immune suppression, such as DMARDS, NSAIDS, corticosteroids, immunosuppressants leading to systemic toxicity, risk of infections, and malignancies [17]. Due to these serious side effects an important adjuvant and complementary therapy is taking an interesting twist in the treatment of RA concerning dietary supplements which involves extraction of polyphenols, a highly water-soluble chemical compound derived from a diverse class of medicinal plants. Polyphenols are shown to have definite clinical efficacy, properties, such as multi-mechanism in pharmacology, multi-component, multi target, no drug-resistance and less toxic side effects making them to be an interesting chemical component to be worked upon [18]. This study primarily focuses on phytochemistry and therapeutic properties of *Nigella sativa* (black seed), whose traditional use as a natural medicine dates back as early as 1st century A.D. [19]. Due to the high levels of various bioactive compounds this plant is considered for its therapeutic effects, antioxidant, anti-inflammatory, anti-allergic, anti-cancer, hypolipidemic and immunomodulating properties [20].

The current study is based on in-silico methodology–molecular docking to evaluate the efficacy of the bioactive compounds using computational tools by analyzing various properties of the compound that are already available in the databases. Therefore, in this research, we have thoroughly analyzed therapeutic properties of bioactive compounds obtained in *Nigella sativa* as a potential inhibitor of inflammatory responses by suppressing the upregulation of NF-KB protein in the treatment of RA.

## MATERIALS AND METHODS

### Preparation and Selection of the Target Protein

The 3D structure of the target protein NF-KBP52 homodimer-DNA complex (1A3Q) was downloaded in PDB format from RCSB Databank (<https://www.rcsb.org/structure/1A3Q>) [21]. Protein was prepared in Discovery Studio 2021 v21. 1. 0. 20298 (BIOVIA) software were the protein is analyzed and cleaned of hetatms, ligands, water molecules, polar hydrogen atoms were added and the protein binding sites were retained [22]. The 3D structure of the altered protein with A and B is then saved in PDB format to proceed ahead with the molecular docking using PYRX.

### Preparation and Selection of the Ligand

30 bioactive compounds from *Nigella sativa* plant were chosen from IMPPAT Database (<https://cb.imsc.res.in/imppat/>) [23]. The canonical smiles and 3D structure data file (SDF) format was downloaded from Pubchem (<https://pubchem.ncbi.nlm.nih.gov/>) [24]. Further analysis of the compounds for physicochemical, pharmacokinetics, drug likeness was done using web tool Swiss ADME (<http://www.swissadme.ch/index.php#1>) [25]. Out of 30 compounds, 18 most favorable compounds were shortlisted based on ADME screening to further determine the potential and binding affinity of the compounds via molecular docking against the target protein.

### In Silico ADME Analysis

During the selection of the ligands, undesirable toxicity and pharmacokinetics of the compounds are to be strictly screened to avoid any adverse effects in the later stages of drug research and development processes. One such tool used to analyze the most desirable compound is Swiss ADME, which analyzes various parameters, such as absorption, distribution, metabolism, excretion, toxicity, pharmacokinetics, and medicinal chemistry, to mention a few. Emphasis on 0 violation Lipinski parameters is taken into consideration while screening and validating the most favorable compounds to carry further for docking

analysis. 30 compounds from *Nigella sativa* were chosen for the ADME analysis. Compounds with high GIA absorption, Lipinski 0 violation filters ( $MW \leq 500$ ,  $MLogP \leq 4$ , 15, N or O  $\leq 10$ , NH or OH  $\leq 5$ ) were strictly followed. 18 compounds passed the above parameters and were carried forward in the research.

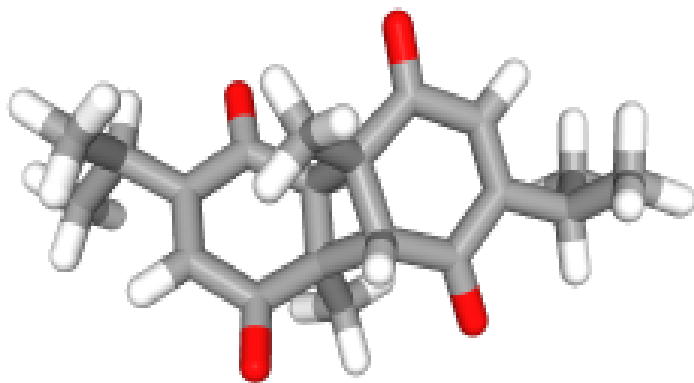
### Molecular Docking

The in-silico target-based screening on the purified protein 1A3Q was carried out in PyRx [26]. The protein was purified in BIOVIA TOOL and was saved in pdbqt format. The PDB file of the prepared protein was loaded, and by autodock, converted into pdbqt format in PyRx. The ligands were loaded in SDF format into PyRx via open babel tool, energy was minimized and then converted into autodock ligand pdbqt format. The ligands and proteins were then selected, moving onto vina vizard, and the grid dimensions were set. The grid dimensions for purified protein 1A3Q for chains A and B were set in vina vizard are as follows: X:16. 7623 Y:61. 4114 Z:0. 6667 dimensions (angstrom) x:82. 8715 Y:89. 3828 Z:93. 8778 and was subjected to docking. The binding affinity scores obtained for ligands against the target protein were then saved in CVS file format. The results were analyzed, and the binding affinity with the lowest score that coordinated with 0 rmsd were preferred for further confirmation and visualization. The pdbqt file of the ligand was saved in PDB format. The pdbqt files of the protein and the selected ligand were then submitted in the biovia tool for visualization to observe the interaction between the target protein against the chosen ligand.

## RESULTS

### Phytochemical Selection

The seeds of the *Nigella sativa* plant are known infamously as cures for any disease except death. The 30 topmost phytochemicals were chosen from the IMPPAT database and screened. Canonical smiles and 3D SDF downloads of the compounds were obtained from PubChem. 18 compounds passed the screening in ADME analysis. The binding affinity of the ligands ranging from  $-3.7$ , i.e., decanoic acid (PubChem ID: 2969) to  $-7.6$ , i.e., nigellone (PubChem ID: 398941). Nigellone (Figure 1) showed the best binding affinity towards the target protein 1A3Q. The ligands scoring a binding affinity of less than  $-6$  were considered for further analysis of, which the compound reflecting the highest binding is carried forward for molecular docking. The binding affinities of the screened compounds.



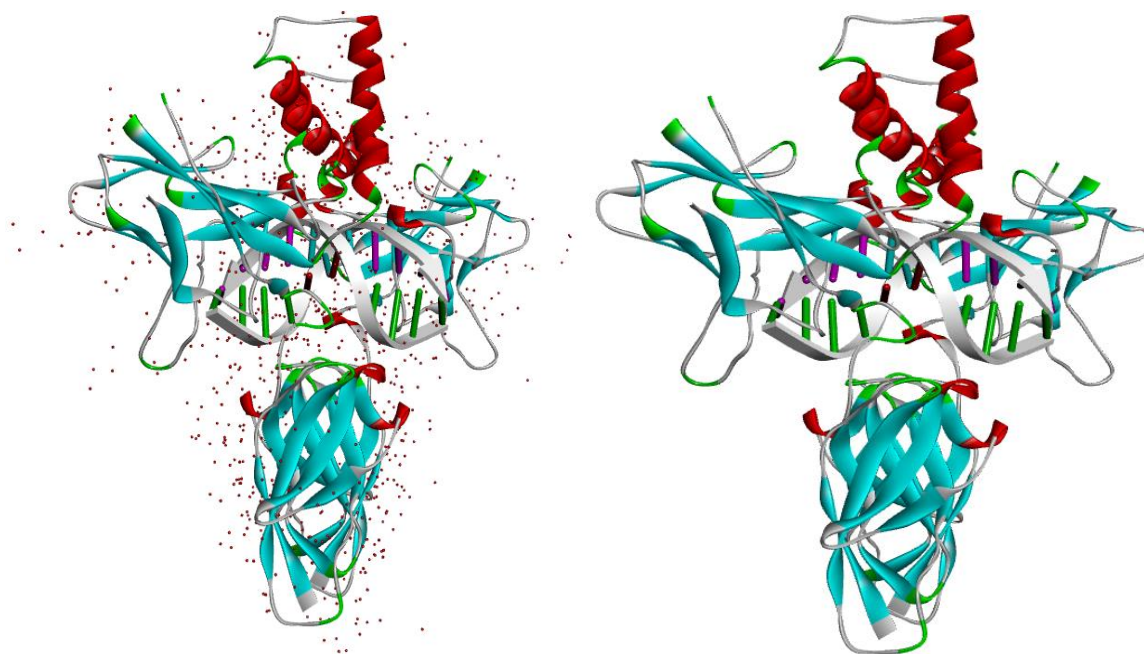
**Figure 1.** 3D structure of the phytochemical nigellone.

### Selection and Purification of the Protein 1A3Q

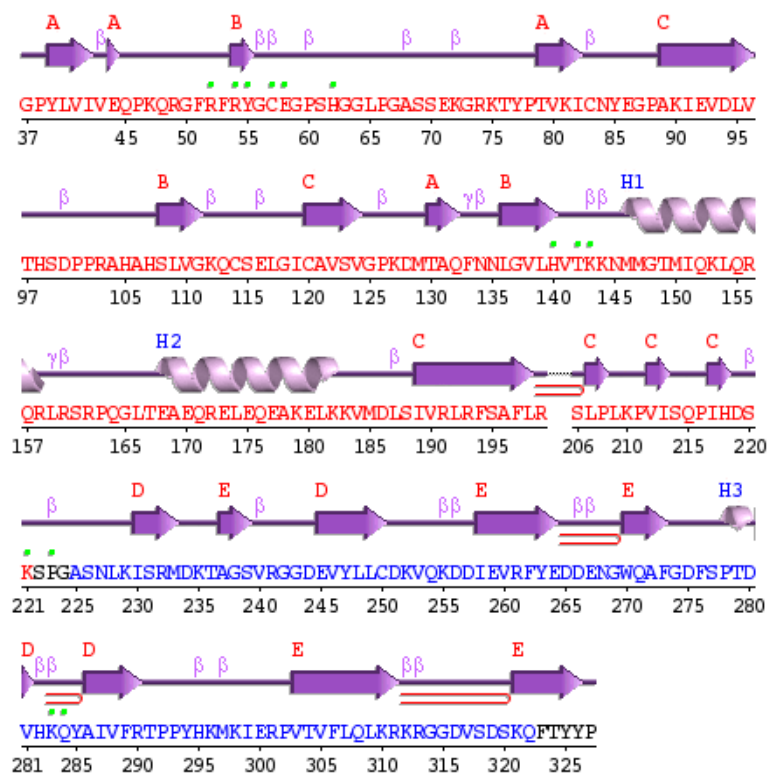
The crystal structure of the human NF-kappaB p52 homodimer-DNA complex obtained through the X-ray diffraction method has a resolution of  $2.1 \text{ \AA}$  with PDB ID-1A3Q and is downloaded in PDB format from the RCSB database (PDB DOI: <https://doi.org/10.2210/pdb1A3Q/pdb>). The protein was then subjected to purification in the BIOVIA tool before the molecular docking process to get the best quality structure where the active sites, A & B chains, and ligand binding sites were restored whilst removing the rest of Figure 2(a, b).

### Structural Analysis of the Protein

The 2D and 3D structural analysis and interaction of the protein 1A3Q was studied in PDBSum (<https://www.ebi.ac.uk/thornton-srv/databases/pdbsum/>) [27]. The protein chains A and B have a total of 285 a. a. residue. The 2D structure of 1A3Q consists of 5 sheets, 4 beta hairpins, 4 beta bulges, 21 strands, 3 helices, 1 helix-helix interaction, 29 beta turns and 2 gamma turn, as shown in Figure 3.



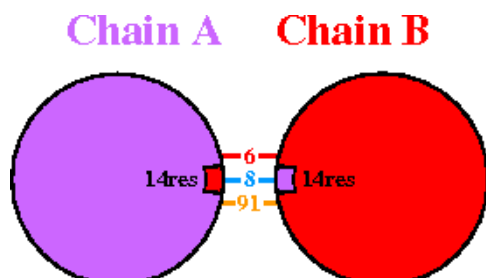
**Figure 2.** 3D Structure of protein 1A3Q. (a) 3D structure from RCSB PDB, (b) Purified 3D structure from BIOVIA.



**Figure 3.** Secondary structure of protein 1A3Q.

### Protein-Protein Interface

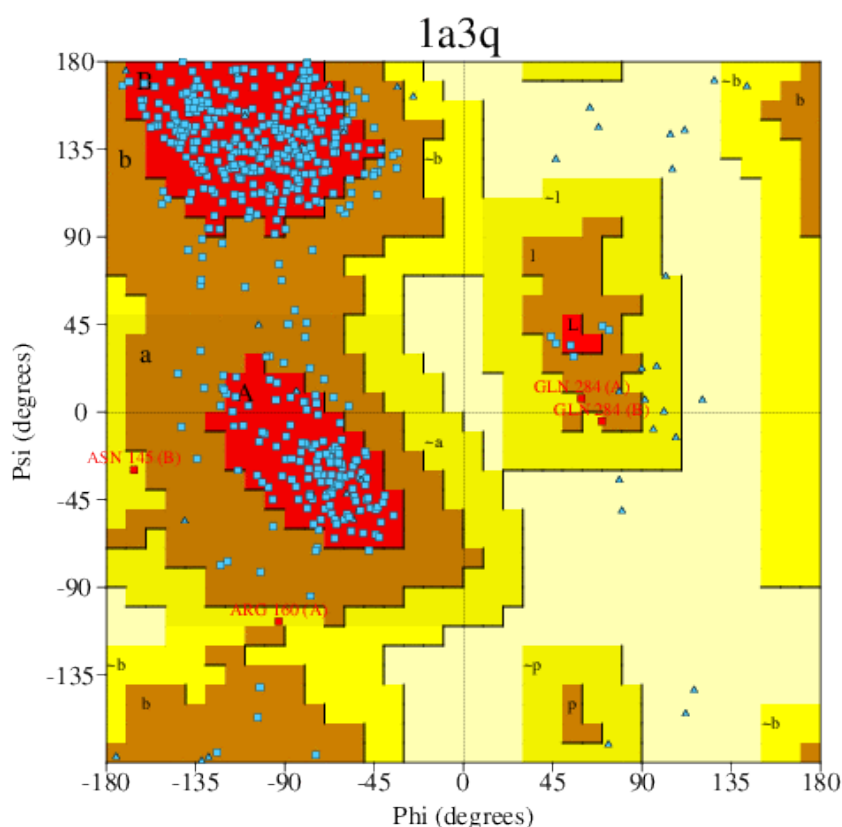
Protein-protein interface statistics were retrieved from PDBsum. Chains A and B have an interface residue of 14 each. Chain A has an interface area ( $A^2$ ) of 691, whereas chain B has an interface area ( $A^2$ ) of 678. Chain A and B were interacting with a total no of 6 salt bridges, 8 Hydrogen bonds, and 91 non-bonded contacts, and there was no evidence of disulphide bonds between the 2 chains (Figure 4).



**Figure 4.** Protein-protein interface of protein 1A3Q from PDBsum.

### Ramachandran Plot

Ramachandran Plot in each protein structure is a display of  $\phi$ ,  $\psi$  angle pairs of a polypeptide chain. The Ramachandran plot was analyzed from PDBSum and PROCHECK as shown in Figure 5. PROCHECK plot statistics observed a total number of 570 residues, of which 482 residues were non-glycine and non-proline. 411 residues (A, B, L) colored red in the plot were classified as more favored regions, 67 residues (a, b, l, p) colored mustard yellow in the additional allowed regions, 4 residues ( $\sim$ a,  $\sim$ b,  $\sim$ l,  $\sim$ p) colored yellow in the additional allowed regions, 0 disallowed regions, and 4 end residues (excluding glycine and proline), of which 46 glycine and 38 proline residues are observed. In in-silico studies, stronger peptide confirmation is observed when the residues fall under the most favorable region.



**Figure 5.** Ramachandran plot of protein 1A3Q.

### Protein Statistics

EMBOSS:pepstats ([https://www.ebi.ac.uk/jdispatcher/seqstats/emboss\\_pepstats](https://www.ebi.ac.uk/jdispatcher/seqstats/emboss_pepstats)) tool analyzes various sequence statistics on the protein properties and this data obtained is crucial for analysis for further protein purification and in R&D [28]. The following details were obtained for protein 1A3Q. molecular weight = 932.07, residues = 11, average residue weight = 84.733, charge = 0.0 isoelectric point = 5.4755. Protein dynamics like number of amino acid residues, their property and % mole were retrieved from EMBOSS pepstats (Table 1).

**Table 1.** 1A3Q Protein statistics data obtained from EMBOSS:pepstats.

Property	Residues	Number	Mole%
Tiny	(A+C+G+S+T)	11	100
Small	(A+B+C+D+G+N+P+S+T+V)	11	100
Aliphatic	(A+I+L+V)	1	9.091
Aromatic	(F+H+W+Y)	0	0
Non-polar	(A+C+F+G+I+L+M+P+V+W+Y)	9	81.818
Polar	(D+E+H+K+N+Q+R+S+T+Z)	2	18.182
Charged	(B+D+E+H+K+R+Z)	0	0
Basic	(H+K+R)	0	0
Acidic	(B+D+E+Z)	0	0

### Pharmacological Studies

The selected 18 ligands of the *Nigella sativa* were subjected to Swiss ADME analysis wherein the phytochemicals were observed under physicochemical parameters, Drug likeness properties pertaining to 0 violation Lipinski rule as shown in Tables 2, 3 & 4. The pharmacokinetics properties of the compounds showed high GI absorption indicating good efficacy, BBB (Blood brain barrier) is shown to be high in the compounds as well, which could be subjected to chemical synthesis to get a desirable composition during drug development considering other beneficial properties in the compounds and less side effects when compared to conventional treatment, Pg-p substrate except for nigellidine is shown to be favorable too.

**Table 2.** Data analyzed for the physicochemical properties from Swiss ADME.

Ligand	Formula	MW	H A (<10)	H D (<5)	TPSA
Myrtenol	C10H16O	152.23	1	1	20.23
Nigellidine	C18H18N2O2	294.35	2	1	47.16
Dillapiol	C12H14O4	222.24	4	0	36.92
Thymoquinone	C10H12O2	164.2	2	0	34.14
Carvacrol	C10H14O	150.22	1	1	20.23
Geranylacetone	C13H22O	194.31	1	0	17.07
Thymol methyl ether	C11H16O	164.24	1	0	9.23
Nigellidine	C13H14N2O3	246.26	3	1	64.23
2-Tridecanone	C13H26O	198.34	1	0	17.07
2-(4-Methylphenyl) propan-2-ol	C10H14O	150.22	1	1	20.23
Lauric acid	C12H24O2	200.32	2	1	37.3
Decanoic acid	C10H20O2	172.26	2	1	37.3
Myristicin	C11H12O3	192.21	3	1	27.69
Nigellone	C20H24O4	328.4	4	0	68.28
Coumarin	C9H6O2	146.14	2	0	30.21
Nonanal	C9H18O	142.24	1	0	17.07
Eugenol	C10H12O2	164.2	2	1	29.46
4-Isopropylbenzaldehyde	C10H12O	148.2	1	0	17.07

Note: MW: molecular weight (<500 daltons), HA: hydrogen acceptor, HD: hydrogen donor, TPSA: topological polar surface area.

**Table 3.** Pharmacokinetics data obtained from Swiss ADME.

Ligand	GI	BBB	P-gp Substrate
Myrtenol	High	Yes	No
Nigellidine	High	Yes	Yes
Dillapiol	High	Yes	No
Thymoquinone	High	Yes	No
Carvacrol	High	Yes	No
Geranylacetone	High	Yes	No
Thymol methyl ether	High	Yes	No
Nigellicine	High	Yes	No
2-Tridecanone	High	Yes	No
2- (4-Methylphenyl) propan-2-ol	High	Yes	No
Lauric acid	High	Yes	No
Decanoic acid	High	Yes	No
Myristicin	High	Yes	No
Nigellone	High	Yes	No
Coumarin	High	Yes	No
Nonanal	High	Yes	No
Eugenol	High	Yes	No
4-Isopropylbenzaldehyde	High	Yes	No

Note: GI: Gastrointestinal absorption, BBB: blood brain barrier, P-gp: permeable-glycoprotein substrate.

**Table 4.** Drug likeness of the phytochemicals obtained from Swiss ADME.

Ligand	Lipinski # Violations	Bioavailability Score
Myrtenol	0	0.55
Nigellidine	0	0.55
Dillapiol	0	0.55
Thymoquinone	0	0.55
Carvacrol	0	0.55
Geranylacetone	0	0.55
Thymol methyl ether	0	0.55
Nigellicine	0	0.85
2-Tridecanone	0	0.55
2- (4-Methylphenyl) propan-2-ol	0	0.55
Lauric acid	0	0.85
Decanoic acid	0	0.85
Myristicin	0	0.55
Nigellone	0	0.55
Coumarin	0	0.55
Nonanal	0	0.55
Eugenol	0	0.55
4-Isopropylbenzaldehyde	0	0.55

### Structural analysis of protein 1A3Q

The NF-Kappa B p52 a homodimer and eukaryotic transcription factor plays a significant role in regulating many mammalian genes encoding pro inflammatory cytokines, major histocompatibility complex (MHC) proteins, cell adhesion molecules, growth factors, interferones including viruses, such as herpes, human immunodeficiency virus (HIV) [29]. 1A3Q human NF-Kappa-B P52 bound to DNA is selected as a macromolecule for this study from RCSB PDB. The 1A3Q protein has 285 sequences

and consists of C [auth A], D [auth B]. Analysis of the protein using prankweb server retrieved 8 active sites. 12 amino acids residues were obtained in pocket 1 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 11 in pocket 2B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 11 in pocket 3 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 9 from pocket 4 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 11 from pocket 5 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 12 from pocket 6 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 10 from pocket 7 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58, 7 from pocket 8 B\_140 B\_142 B\_143 B\_187 B\_188 B\_219 B\_221 B\_222 B\_223 B\_55 B\_57 B\_58 [30]. No mutagenesis was reported on protein 1A3Q explored with computed atlas of surface topography of proteins (CASTp) ([http://sts.bioe.uic.edu/castp/index.html?j\\_65e41d83b2dbf](http://sts.bioe.uic.edu/castp/index.html?j_65e41d83b2dbf)) [31].

### Molecular Docking Analysis

Prepared protein 1A3Q and the ligands were docked in PyRx. Docking results were obtained from PyRx and the results were visualized in BIOVIA (Table 5). Receptor protein and the ligand were defined and the 2D visualization confirmed the interaction of the selected ligand with amino acids in the protein by conventional hydrogen bond and Pi-alkyl bond. The binding affinity of the 18 ligands with respect to A and B chains of the proteins are as shown in Table 6. Binding score less than -6 were considered for further analysis and the ligands nigellone (398941), nigellidine (138628302), nigellicine (11402337) as referred in Table 1 showed favorable binding affinity of which the highest binding affinity observed in nigellone was carried forward for further visualization in BIOVIA.

### Protein Rretrieved from RCSB PDB

#### *Binding Affinity of Nigella Sativa Phytocompounds with 1A3Q*

18 compounds were screened and the binding affinity of less than -6 was favored. 3 compounds that displayed binding affinity towards the target protein are as shown in Table7. Nigellone with binding affinity of -7.6 was docked and visualized against the target protein in BIOVIA. Confirmation of interactions in 2D of the docked complex can be observed in Figure 6.

**Table 5.** 1A3Q protein details obtained from RCSB PDB.

Molecule	Chain	Sequence Length	Organism	Details
NF-KAPPA B PROTEIN	C [auth A], D [auth B]	285	<i>Homo sapiens</i>	Transcription/DNA Mutation (s) -NO

### 2D Visualization and Molecular Interactions of 1A3Q Protein with Ligand Nigellone

Amino acids ARG B-193, ARG B-103, ARG B-103, and ARG B-160 (Table 8) is shown to have strong conventional hydrogen bond interaction and amino acid PHE B-197 is interacting with pi-alkyl bond with the ligand nigellone (398941) obtaining a binding affinity of -7.6 with respect to B chains of 1A3Q protein as shown in Figure 6.

## DISCUSSION

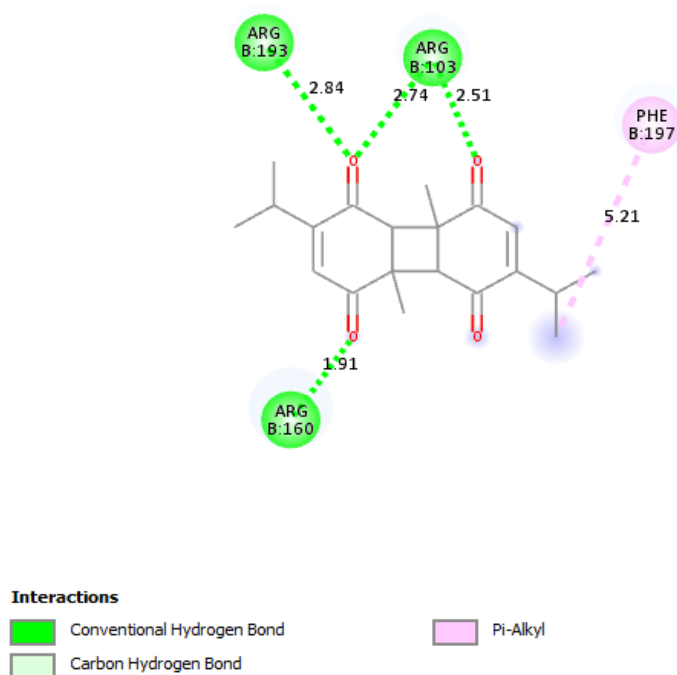
RA is a chronic inflammatory progressive autoimmune disease that gradually affects the synovial joints and leads to subsequent erosion and deformity of bone and joints. The true etiology of this disease is complex and multifactorial and certain conditions like environmental factors and genetics play a crucial role in outset and predisposition of this disease. RA manifests in either of 2 states active or inactive, while in active state tissues are inflamed and the relapse symptoms flares up, in an inactive state the inflammation subsides [3, 32, 33]. RA linked to environmental factors has gained attention in recent years from epidemiological and animal studies for their apparent role in provoking autoimmunity in an already genetically predisposed individual [34, 35]. These factors have shown to increase the oxidative stress in an individual creating a negative balance in the pool of oxidative molecules favoring predominance of the prooxidants leading to elevated concentrations of ROS, RNS, lipid peroxidation damaging the cellular components which are directly involved in the pathogenesis of RA [10, 36–37].

**Table 6.** A and B chain binding affinity data of protein 1A3Q with Phytocompounds of *Nigella sativa* from PyRx.

Ligand	Binding Affinity
Myrtenol	-5.9
Nigellidine	-6.9
Dillapiol	-5.2
Thymoquinone	-5.4
Carvacrol	-5.4
Geranylacetone	-4.8
Thymol methyl ether	-5.1
Nigellicine	-6.4
2-Tridecanone	-4
2-(4-Methylphenyl) propan-2-ol	-5.4
Lauric acid	-4.3
Decanoic acid	-3.7
Myristicin	-4.8
Nigellone	-7.6
Coumarin	-5.7
Nonanal	-4.2
Eugenol	-4.8
4-Isopropylbenzaldehyde	-5

**Table 7.** Binding affinity of top 3 phytocompounds with protein 1A3Q.

Phytocompound Name	PubChem ID	Binding Affinity in kcal/mol
Nigellone	398941	-7.6
Nigellidine	138628302	-6.8
Nigellicine	11402337	-6.4



**Figure 6.** 2D visualization of nigellone interacting with protein 1A3Q.

**Table 8.** Protein complex 1A3Q and nigellone docking interactions.

Name	Distance	Category	Types
ARG B-193	2.84	Hydrogen bond	Conventional hydrogen bond.
ARG B-103	2.74	Hydrogen bond	Conventional hydrogen bond.
ARG B-103	2.51	Hydrogen bond	Conventional hydrogen bond.
ARG B-160	1.91	Hydrogen bond	Conventional hydrogen bond.
PHE B-197	5.21	Hydrophobic	Pi-alkyl.

Cytoplasmic eukaryotic transcription factor NF- $\kappa$ B was shown to be directly influenced by oxidative stress and takes a key role in regulating the genes involved in immune and inflammatory processes [38–41]. NF- $\kappa$ B signals can either be triggered or suppressed by free radicals and are variably context dependent and induce pro and antioxidant role in inducing oxidative stress. The upregulation of NF- $\kappa$ B target genes is led by 2 signaling pathways described as canonical (classic pathway) and non-canonical (alternative pathway). In RA NF- $\kappa$ B p52 is involved in inducing inflammation through non-canonical pathway that primarily activate the TNF receptors triggered by lymphotoxin  $\beta$  receptor, CD40 and B cell activation factor receptor 3 usually in concurrence with RelA by cytokine receptors (IL-1R, TNFR) [42–44]. NF- $\kappa$ B plays a key role in inducing inflammatory pathways in RA related to oxidative stress induced by free radicals, working on inhibiting free radicals through powerful antioxidants by reducing oxidative stress which in turn would downregulate the activity of NF- $\kappa$ B and hence reducing the inflammation is the primary principle involved in targeting NF- $\kappa$ B as a protein of choice in this study. The current treatment plan that has proven to suppress inflammation on targeting free radicals and has shown subsequent reduction in pannus proliferation is NOS inhibitor L-NAME GW274150 [45, 46]. Among the other treatment plans, first-line management: NSAIDs and corticosteroids, opioid analgesics, second-line management: disease-modifying antirheumatic drugs (DMARD) taking part as immunosuppressants are directed towards reducing the inflammation however the adverse side effects like high liver toxicity, bone marrow deterioration, cirrhosis, hypertension, leukopenia, GI upset, interstitial lung disease are some of the drawbacks that are accompanied along with the treatment [47].

Pharmaceutical industries have always been attracted towards plant-based drugs for their convenience in standardization and purification of the compounds while being more efficacious with minimum side effects, by large harmless, easy metabolism and absorption in the body when compared to synthetic medicine [48]. For many centuries medicinal plants have been used in herbal medicines for their abundance in two major antioxidant phytochemicals carotenoids and polyphenols. *Nigella sativa* (*N. sativa*) (family *Ranunculaceae*) commonly known as black seed is well known for its wide spectrum of pharmacological potential as an anticancer and immunomodulatory, antimicrobial, analgesic and anti-inflammatory, gastroprotective, renal protective, hepatoprotective, anthelmintics, diuretic, antidiabetic, and antioxidant properties [49]. In this study we have chosen *Nigella sativa* as a potential drug supplement and as alternative therapy for allopathic medicine in RA to treat the free radical formation by high antioxidant properties of the phytochemicals present in *N. sativa*. In vitro studies indicate the bioactive compound thymoquinone at a dose 1–5  $\mu$ M can effectively cause apoptosis and inhibit IL-6 and IL-8 induced by TNF- $\alpha$  by blocking the expression of myeloid leukemia (Mcl) [15]. Experiments done by Arjumand et al. have reported bioactive compound thymoquinone (TQ, 5-isopropyl-2-methyl-1, 4-benzoquinone) treatment conducted on 6–8 weeks old 40 male rats sprague-dawley with a dosage of 10 mg/kg body weight for 20 days showed reduced macroscopic arthritis score, CRP levels, synovitis, bone erosion and pannus formation, downregulation of NF- $\kappa$ B mRNA, decreased levels of TLR2, TLR4, IL-1 and TNF- $\alpha$  and showed no signs of hepatotoxicity or nephrotoxicity [15, 50]. Clinical trials on humans by Hadi et al. prescribed with black cumin seed oil capsules 2 capsules, 500 mg/day (1 g/day) for 8 weeks, on a study group of 50 patients 16-placebo group, 23 intervention group with 39 complete completion showed significant decrease in intervention group in the DAS-28 compared to placebo group, reduction in MDA and NO to baseline value ( $p < 0.05$ ) comparison in the serum levels, increased serum IL-10 levels ( $p < 0.01$ ) in intervention group. There was no significant difference in serum levels of TNF- $\alpha$ , MDA, catalase, IL-10 and TAC among intervention and placebo group [15]. In

silico molecular docking analysis on the bioactive compounds in *N. sativa* observed nigellone (PubChem ID-398941) to be the most bioactive compound having the highest binding affinity against the target protein 1A3Q followed by nigellidine and nigellicine shown in Table 1, whereas thymoquinone (PubChem ID-10281) showed a binding affinity of  $-5.4$ . The current study has proven nigellone to be the most preferred compound when targeted against the protein NF-KB at a molecular level. ADMETLab2.0 toxicity on the liver DILI was shown to be high in the compound, which is also the main side effect observed in the current DMARDS methotrexate treatment. Insight into this concern can be taken into consideration during the drug design and development process to chemically synthesize the compound to a lesser toxic level and this should be able to outweigh the limitation when compared to the potentiality of the compound. Therefore, further in vitro and in vivo studies on nigellone may show better inhibition of NF-KB thus alleviating inflammation and free radical generation in the patients and could be considered as an antioxidant dietary supplement gradually increasing the dosage alongside conventional treatment with minimal side effects.

Previous studies conducted on *Nigella sativa* in vitro and in-vivo have shown thymoquinone to be the highly potent among the phytochemicals, however in-silico analysis of the phytochemicals targeted against NF-KB protein responsible in perpetuating inflammatory responses revealed nigellone (dithymoquinone) to have stronger binding affinity with the protein as discussed previously hypothesizing the emphasis on the compound for extensive research. Validation of the experimental results can further be concluded by dynamic simulation to capture the interactions, movement, functioning and behavior of protein 1A3Q at an atomic level with nigellone (dithymoquinone) in action over time and could be considered as a novel compound to carry forward in future studies in suppressing the inflammation in RA caused by free radicals.

## CONCLUSIONS

Free radicals formed due to decrease in antioxidant levels in the body leading to oxidative stress plays a major role in the pathogenesis of RA by upstreaming the activity of NF-KB causing DNA damage. Raising the antioxidant levels in the body and by negating the proliferation of free radicals could be a propitious strategy in decreasing the inflammation in RA patients. *Nigella sativa*'s highly potent antioxidant properties evidenced for centuries together and believed to be a miracle herb across civilizations marked and proven by scientific studies for its healing traits could turn out to be revolutionary as an antioxidant drug, which is the root cause for escalating pathological conditions. Previous experimental studies have shown desirable results with respect to thymoquinone as the most effective phytochemical. The current molecular docking interactions have unveiled nigellone (dithymoquinone) to be exhibiting the highest binding affinity with NF-KB, however data specific to nigellone is currently scarce to support this study. On the positive end due to the similarities between the structures of the phytochemicals, nigellone can be considered as a stronger alternative prospect expected to bring forth better results in the future research studies.

## Abbreviations

GI	:	Gastro intestinal
BBB	:	Blood brain barrier
P-gp	:	Permeable-glycoprotein substrate
OS	:	Oxidative stress
DMARD	:	Disease modifying anti-rheumatic drug
NSAIDS	:	Non-steroidal anti-rheumatic drugs
P-gp	:	Permeable-glycoprotein substrate
RNS	:	Reactive nitrogen species
TNF	:	Tumor necrosis factor
CD40	:	Cluster of differentiation 40
IL-1R	:	Interleukin 1 receptor
NOS	:	Nitric oxide synthase

---

L-NAME	:	Levo-nitroarginine methyl ester
TLR	:	Toll like a receptor
MDA	:	Malondialdehyde
TAC	:	Total antioxidant capacity
DILI	:	Drug induced liver injury

### Acknowledgment

The author hereby acknowledges the Department of BioNome, Bengaluru, India (<https://bionome.in/>) for providing computation facilities and assistance in conducting this research. She thanks Ms. Samiksha Bhor for her guidance and supervision throughout the project.

### Funding

Not applicable.

### Authors' Contribution

All the authors have equally contributed to the manuscript.

### Conflict of Interest

The authors declare no conflict of interest.

### REFERENCES

1. Ferreira HB, Melo T, Paiva A, Domingues MDR. Insights in the role of lipids, oxidative stress and inflammation in rheumatoid arthritis unveiled by new trends in lipidomic investigations. *Antioxidants*. 2021;10(1):45. doi:10.3390/antiox10010045.
2. Black RJ, Cross M, Haile LM, Culbreth GT, Steinmetz JD, Hagins H, et al. Global, regional, and national burden of rheumatoid arthritis, 1990–2020, and projections to 2050: a systematic analysis of the Global Burden of Disease Study 2021. *Lancet Rheumatol*. 2023;5(10):e594–e610. doi:10.1016/S2665-9913(23)00211-4.
3. Jahid M, Khan KU, Rehan-Ul-Haq, Ahmed RS. Overview of rheumatoid arthritis and scientific understanding of the disease. *Mediterr J Rheumatol*. 2024;34(3):284.
4. Cross M, Smith E, Hoy D, Carmona L, Wolfe F, Vos T, et al. The global burden of rheumatoid arthritis: estimates from the global burden of disease 2010 study. *Ann Rheum Dis*. 2014;73(7):1316–1322. doi:10.1136/annrheumdis-2013-204627.
5. Hunter TM, Boytsov NN, Zhang X, Schroeder K, Michaud K, Araujo AB. Prevalence of rheumatoid arthritis in the United States adult population in healthcare claims databases, 2004–2014. *Rheumatol Int*. 2017;37(9):1551–1557. doi:10.1007/s00296-017-3726-1.
6. Tanner S, Dufault B, Smolik I, Meng X, Anaparti V, Hitchon C, et al. A prospective study of the development of inflammatory arthritis in the family members of Indigenous North American people with rheumatoid arthritis. *Arthritis Rheumatol*. 2019;71(9):1494–1503. doi:10.1002/art.40880.
7. du Montcel ST, Michou L, Petit-Teixeira E, Osorio J, Lemaire I, Lasbleiz S, et al. New classification of HLA–DRB1 alleles supports the shared epitope hypothesis of rheumatoid arthritis susceptibility. *Arthritis Rheumatol*. 2005;52(4):1063–1068. doi:10.1002/art.20989.
8. El-Gabalawy HS, Maisha JA, O’Neil LJ. Modifiable risk factors linked to the development of rheumatoid arthritis: evidence, immunological mechanisms and prevention. *Front Immunol*. 2023;14. doi:10.3389/fimmu.2023.1221125.
9. Wang X, Fan D, Cao X, Ye Q, Wang Q, Zhang M, et al. The role of reactive oxygen species in the rheumatoid arthritis-associated synovial microenvironment. *Antioxidants*. 2022;11(6):1153. doi:10.3390/antiox11061153.
10. Fonseca LJS da, Nunes-Souza V, Goulart MOF, Rabelo LA. Oxidative stress in rheumatoid arthritis: What the future might hold regarding novel biomarkers and add-on therapies. *Oxid Med Cell Longev*. 2019;2019(1). doi:10.1155/2019/7536805.
11. Sehnert B, Burkhardt H, Dübel S, Voll RE. Cell-type targeted NF-kappaB inhibition for the treatment of inflammatory diseases. *Cells*. 2020;9(7):1627. doi:10.3390/cells9071627.

12. Jimi E, Huang F, Nakatomi C. NF- $\kappa$ B signaling regulates physiological and pathological chondrogenesis. *Int J Mol Sci.* 2019;20(24):6275. doi:10.3390/ijms20246275.
13. Zielińska M, Dereń K, Polak-Szczybyło E, Stępień AE. The role of bioactive compounds of *Nigella sativa* in rheumatoid arthritis therapy—current reports. *Nutrients.* 2021;13(10):3369. doi:10.3390/nu13103369.
14. Tekeoglu I, Dogan A, Demiralp L. Retracted: Effects of thymoquinone (volatile oil of black cumin) on rheumatoid arthritis in rat models. *Phytother Res.* 2007;21(9):895–897. doi:10.1002/ptr.2143.
15. Hadi V, Naseh P, Mahsa M, Elyas NE, Gholizadeh NJ, Saeid H. *Nigella sativa* in controlling type 2 diabetes, cardiovascular, and rheumatoid arthritis diseases molecular aspects. *J Res Med Sci.* 2021;26(1):20. doi:10.4103/jrms.JRMS\_236\_20.
16. Radu AF, Bungau SG. Management of rheumatoid arthritis: An overview. *Cells.* 2021;10(11):2587. doi:10.3390/cells10112857.
17. Long Z, Xiang W, He Q, Xiao W, Wei H, Li H, et al. Efficacy and safety of dietary polyphenols in rheumatoid arthritis: A systematic review and meta-analysis of 47 randomized controlled trials. *Front Immunol.* 2023;14. doi:10.3389/fimmu.2023.1024120.
18. Salehi B, Quispe C, Imran M, Iahtisham-Ul-Haq, Zivkovic J, Abu-Reidah IM, et al. *Nigella* plants—traditional uses, bioactive phytoconstituents, preclinical and clinical studies. *Front Pharmacol.* 2021;12. doi:10.3389/fphar.2021.625386.
19. Dalli M, Bekkouch O, Azizi SE, Azghar A, Gseyra N, Kim B. *Nigella sativa* L. phytochemistry and pharmacological activities: a review (2019–2021). *Biomol.* 2022;12(1):20. doi:10.3390/biom12010020.
20. Berman HM, Battistuz T, Bhat TN, Bluhm WF, Bourne PE, Burkhard K, et al. The Protein Data Bank. *Nucleic Acids Res.* 2000;28:235–242. doi:10.1107/S0907444902003451.
21. Kemmish H, Fasnacht M, Yan L. Fully automated antibody structure prediction using BIOVIA tools: Validation study. *PLoS One.* 2017;12(5):e0177923. doi:10.1371/journal.pone.0177923.
22. Mohanraj K, Karthikeyan BS, Vivek-Ananth RP, Chand RPB, Aparna SR, Mangalapandi P, et al. IMPPAT: A curated database of Indian medicinal plants, phytochemistry and therapeutics. *Sci Rep.* 2018;8(1):4329. doi:10.1038/s41598-018-22631-z.
23. Kim S, Thiessen PA, Bolton EE, Chen J, Fu G, Gindulyte A, Han L, He J, He S, et al. PubChem substance and compound databases. *Nucleic Acids Res.* 2016;44(D1):D1202–D1213. doi:10.1093/nar/gkv951.
24. Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Sci Rep.* 2017;7:42717. doi:10.1038/srep42717.
25. Dallakyan S, Olson AJ. Small-Molecule Library Screening by Docking with PyRx. In: Hempel J, Williams C, Hong C, editors. *Chemical Biology. Methods in Molecular Biology.* New York, NY: Humana Press; 2015. 243–250. doi:10.1007/978-1-4939-2269-7\_19.
26. Laskowski RA, Jabłońska J, Pravda L, Vařeková RS, PDBsum JT. Structural summaries of PDB entries. 2018;27. doi:10.1002/pro.;3289:129-34.
27. <http://cib.cf.ocha.ac.jp/bitool/EMBOSS-6.0.1/doc/programs/master/emboss/apps/pepstats.html>.
28. Larson CJ, Verdine GL, Müller CW. Structure of the human NF-kappaB p52 homodimer-DNA complex at 2.1 Å resolution. *EMBO J.* 1997;16:7078–7090. doi:10.1093/emboj/16.23.7078.
29. Jendele L, Krivak R, Skoda P, Novotny M, Hoksza D. PrankWeb: a web server for ligand binding site prediction and visualization. *Nucleic Acids Res.* 2019;47(W1):W345–349. doi:10.1093/nar/gky424.
30. Tian W, Chen C, Lei X, Zhao J, CAST JL. 3.0: Computed atlas of surface topography of proteins. 2018;46:W363–W367. doi:10.1093/nar/gky473.
31. Venables P, Maini RN. Diagnosis and differential diagnosis of rheumatoid arthritis. *UpToDate2012.* 2014.
32. Rossetti M, Spreafico R, Consolaro A, Leong JY, Chua C, Massa M, et al. TCR repertoire sequencing identifies synovial treg cell clonotypes in the bloodstream during active inflammation in human arthritis. *Ann Rheum Dis.* 2017;76(2):435–441. doi:10.1136/annrheumdis-2015-208992.

33. Long H, Yin H, Wang L, Gershwin ME, Lu Q. The critical role of epigenetics in systemic lupus erythematosus and autoimmunity. *J Autoimmun.* 2016;74:118–138. doi:10.1016/j.jaut.2016.06.020.
34. Khan MF, Wang G. Environmental agents, oxidative stress and autoimmunity. *Curr Opin Toxicol.* 2018;7:22–27. doi:10.1016/j.cotox.2017.10.012.
35. Smallwood MJ, Nissim A, Knight AR, Whiteman M, Haigh R, Winyard PG. Oxidative stress in autoimmune rheumatic diseases. *Free Radic Biol Med.* 2018;125:3–14. doi:10.1016/j.freeradbiomed.2018.05.086
36. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev.* 2002;82(1):47–95. doi:10.1152/physrev.00018.2001.
37. Schreck R, Rieber P, Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1. *EMBO J.* 1991;10(8):2247–2258. doi:10.1002/j.1460-2075.1991.tb07761.x.
38. Rhee SG. Redox signaling: hydrogen peroxide as intracellular messenger. *Exp Mol Med.* 1999;31(2):53–59. doi:10.1038/emm.1999.9.
39. Kondo N, Kanai T, Okada M. Rheumatoid arthritis and reactive oxygen species: a review. *Curr Issues Mol Biol.* 2023;45(4):3000–3015. doi:10.3390/cimb45040197.
40. Makarov SS. NF-kappaB in rheumatoid arthritis: a pivotal regulator of inflammation, hyperplasia, and tissue destruction. *Arthritis Res Ther.* 2001;3:200. doi:10.1186/ar300.
41. Mitchell S, Vargas J, Hoffmann A. Signaling via the NFκB system: signaling via the NFκB system. *Wiley Interdiscip Rev Syst Biol Med.* 2016;8(3):227–241. doi:10.1002/wsbm.1331.
42. Hayden MS, Ghosh S. Shared principles in NF-kappaB signaling. *Cell.* 2008;132(3):344–362. doi:10.1016/j.cell.2008.01.020.
43. Lingappan K. NF-κB in oxidative stress. *Curr Opin Toxicol.* 2018;7:81–86. doi:10.1016/j.cotox.2017.11.002.
44. Seymour M, Pétavy F, Chiesa F, Perry H, Lukey PT, Binks M, et al. Ultrasonographic measures of synovitis in an early phase clinical trial: a double-blind, randomised, placebo and comparator controlled phase IIa trial of GW274150 (a selective inducible nitric oxide synthase inhibitor) in rheumatoid arthritis. *Clin Exp Rheumatol.* 2012;30(2):254–261.
45. Dey S, Bishayi B. Effect of iNOS inhibitor LNMMA along with antibiotics chloramphenicol or ofloxacin in murine peritoneal macrophages regulates *S. aureus* infection as well as inflammation: An in vitro study. *Microb Pathog.* 2017;105:307–320. doi:10.1016/j.micpath.2017.02.031.
46. Bullock J, Rizvi SAA, Saleh AM, Ahmed SS, Ansari RA, Ahmed J, Rheumatoid arthritis: a brief overview of the treatment. *Med Princ Pract.* 2019;27(6):501–507. doi:10.1159/000493390.
47. Nasim N, Sandeep IS, Mohanty S. Plant-derived natural products for drug discovery: current approaches and prospects. 2022;65:399–411. doi:10.1007/s13237-022-00405-3.
48. Ahmad A, Husain A, Mujeeb M, Khan SA, Najmi AK, Siddique NA, Damanhoury ZA, Anwar F. A review on therapeutic potential of *Nigella sativa*: A miracle herb. *Asian Pac J Trop Biomed.* 2013;3(5):337–352. doi:10.1016/S2221-1691(13)60075-1.
49. Arjumand S, Shahzad M, Shabbir A, Yousaf MZ. Thymoquinone attenuates rheumatoid arthritis by downregulating TLR2, TLR4, TNF-α, IL-1, and NFκB expression levels. *Biomed Pharmacother.* 2019;111:958–963. doi:10.1016/j.biopha.2019.01.006.
50. Hadi V, Kheirouri S, Alizadeh M, Khabbazi A, Hosseini H. Effects of *Nigella sativa* oil extract on inflammatory cytokine response and oxidative stress status in patients with rheumatoid arthritis: A randomized; double-blind; placebo-controlled clinical trial. *Avicenna J Phytomed.* 2016;6(1):34–43.