



Genomic Characterization of Emerging Arboviruses in Rural India

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Abstract

Arboviruses (arthropod-borne viruses) represent a rapidly evolving group of pathogens responsible for significant morbidity and mortality, particularly in tropical and subtropical regions. Rural India, characterized by dense vector populations, changing ecological patterns, and limited healthcare infrastructure, has become a hotspot for the emergence and re-emergence of arboviral diseases such as dengue, chikungunya, Japanese encephalitis, and more recently, Zika virus infections. Advances in genomic technologies, including next-generation sequencing (NGS), metagenomics, and phylogenetic analysis, have revolutionized the detection, surveillance, and characterization of these viruses. This review focuses on the genomic characterization of emerging arboviruses in rural India, highlighting their molecular diversity, evolutionary dynamics, and epidemiological implications. It discusses the role of environmental, socio-economic, and ecological factors in viral emergence, and emphasizes the importance of genomic surveillance in early detection and outbreak management. Furthermore, the article explores current challenges, including limited diagnostic infrastructure and gaps in genomic data from rural settings, while proposing future strategies integrating genomics with public health frameworks. The study aims to provide a comprehensive understanding of arboviral genomics in rural India and its relevance to drug development, vaccine design, and novel therapeutic interventions.

Keywords: Arboviruses, genomic characterization, rural India, next-generation sequencing, viral evolution, dengue virus, zika virus, chikungunya, phylogenetics, public health surveillance

INTRODUCTION: OVERVIEW OF ARBOVIRUSES AND THEIR GLOBAL SIGNIFICANCE

Arboviruses, or arthropod-borne viruses, are transmitted to humans and animals through vectors such as mosquitoes, ticks, and sandflies. These viruses belong to diverse viral families, including *Flaviviridae*, *Togaviridae*, and *Bunyaviridae*, and are responsible for a wide spectrum of diseases ranging from mild febrile illness to severe neurological and hemorrhagic conditions [1].

Globally, arboviral infections have gained increasing attention due to their expanding geographic distribution, frequent outbreaks, and potential for epidemic spread. Climate change, globalization, urbanization, and ecological disruptions have significantly contributed to the proliferation of vectors and the emergence of new viral strains (Figure 1).

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Transmission Cycle of Arboviruses

In India, arboviral diseases pose a major public health challenge, particularly in rural regions where environmental conditions favor vector breeding. Poor sanitation, stagnant water bodies, agricultural practices, and close human–animal interactions create ideal conditions for virus transmission. Additionally, limited access to healthcare and diagnostic facilities often leads to underreporting and delayed outbreak response [2].

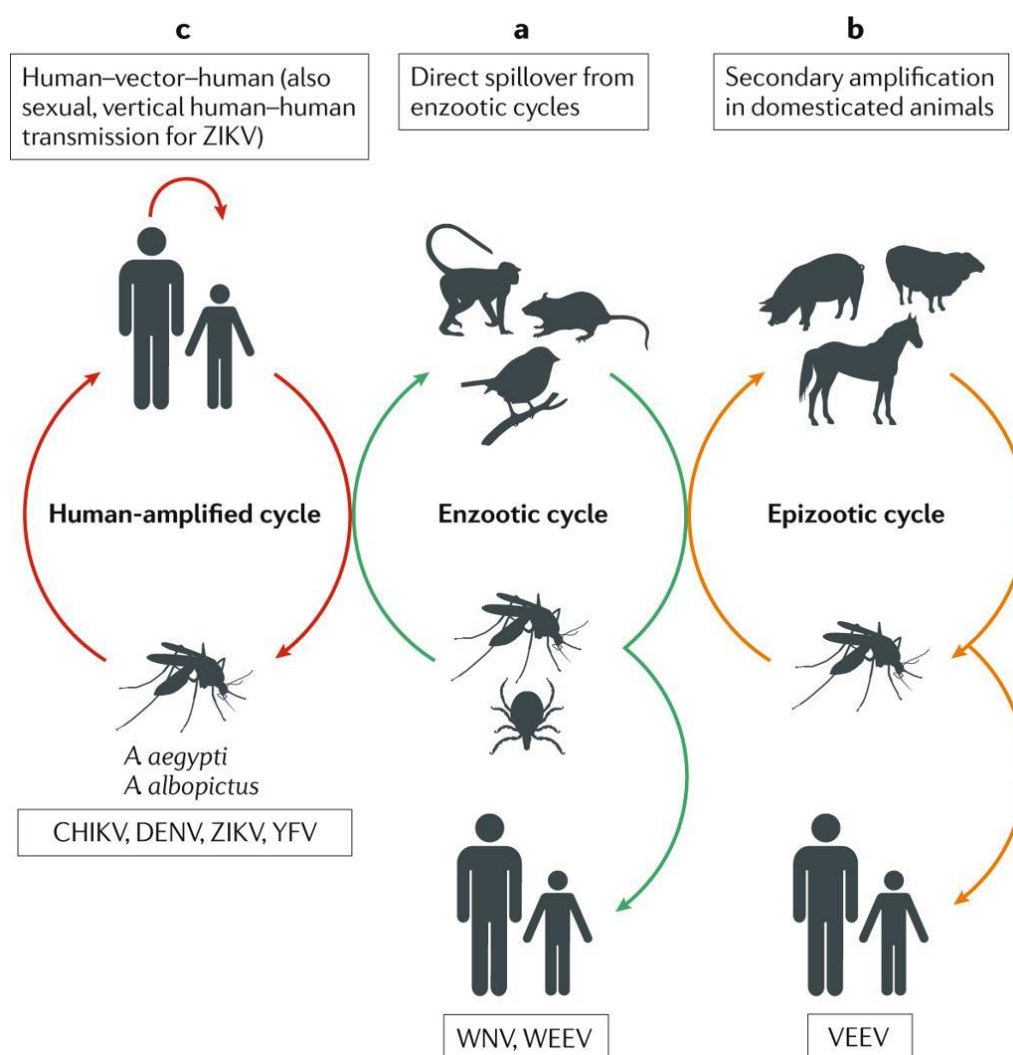


Figure 1. illustrates the complex transmission cycle involving vectors, animal reservoirs, and human hosts, highlighting the zoonotic nature of arboviral infections.

Recent years have witnessed the emergence of novel arboviruses and the re-emergence of previously controlled ones. For instance, dengue has transitioned from an urban-centric disease to a rural endemic problem, while Japanese encephalitis continues to affect children in agricultural communities. The introduction of Zika virus further underscores the dynamic and unpredictable nature of arboviral spread (Table 1).

Table 1. Major arboviruses reported in India.

Virus name	Family	Vector	Disease manifestation	Rural impact
Dengue Virus	Flaviviridae	<i>Aedes aegypti</i>	Dengue fever, hemorrhagic fever	High.
Chikungunya Virus	Togaviridae	<i>Aedes albopictus</i>	Joint pain, fever	Moderate.
Japanese Encephalitis	Flaviviridae	<i>Culex</i> species	Encephalitis	Very High.
Zika Virus	Flaviviridae	<i>Aedes aegypti</i>	Congenital defects, mild fever	Emerging.
Kyasanur Forest Disease Virus	Flaviviridae	Ticks	Hemorrhagic fever	Localized.

The increasing burden of arboviral infections necessitates advanced approaches for detection and characterization. Traditional diagnostic methods, such as serology and PCR, although useful, are limited in their ability to detect novel variants or co-infections. In contrast, genomic characterization provides detailed insights into viral evolution, mutation patterns, and transmission pathways.

Understanding the genomic architecture of arboviruses is crucial for several reasons:

- It aids in identifying virulence factors and drug targets.
- It supports vaccine development through antigenic mapping.
- It enables tracking of viral spread and outbreak origins.
- It helps detect emerging strains with pandemic potential.

In rural India, however, genomic studies remain sparse due to infrastructural and logistical challenges. Bridging this gap is essential for strengthening disease surveillance and improving public health outcomes.

In addition to ecological factors, socio-economic determinants also play a critical role. Limited awareness about vector control, inadequate sanitation, and poor access to preventive healthcare contribute to sustained transmission cycles. Furthermore, migration of laborers between urban and rural areas facilitates the introduction of new viral strains into previously unaffected regions [3].

Another important aspect is the genetic adaptability of both the virus and the vector. Mutations in viral genomes can enhance infectivity, transmission efficiency, and immune evasion. For example, specific mutations in the envelope protein of dengue virus have been associated with increased virulence. Similarly, *Aedes albopictus*, originally a forest-dwelling mosquito, has adapted to rural and urban environments, expanding the geographic reach of chikungunya virus (Table 2).

Table 2. Key drivers of arboviral emergence in rural India.

Factor type	Specific drivers	Impact on arbovirus spread
Environmental	Climate change, rainfall, temperature	Increased vector breeding.
Ecological	Deforestation, wildlife interaction	Zoonotic spillover.
Socio-economic	Poor sanitation, lack of awareness	Sustained transmission.
Biological	Viral mutations, vector adaptability	Enhanced infectivity.
Healthcare-related	Limited diagnostics, underreporting	Delayed outbreak detection.

BACKGROUND: EVOLUTION AND DIVERSITY OF ARBOVIRUSES

Arboviruses exhibit remarkable genetic diversity, which is a key factor contributing to their adaptability and persistence in varied ecological settings. This diversity arises primarily due to the high mutation rates associated with RNA viruses, lack of proofreading mechanisms in RNA-dependent RNA polymerases, and frequent recombination events. Such genetic variability enables arboviruses to evade host immune responses, adapt to new vectors, and expand their host range [4].

Among the major arboviral families, *Flaviviridae* (e.g., dengue, Zika, Japanese encephalitis), *Togaviridae* (e.g., chikungunya), and *Bunyaviridae* (e.g., Crimean–Congo hemorrhagic fever virus) have been extensively studied for their genomic plasticity. The genomes of these viruses are typically single-stranded RNA, either positive-sense or negative-sense, encoding structural and non-structural proteins essential for viral replication and pathogenesis.

The dengue virus (DENV), for instance, exists as four distinct serotypes (DENV-1 to DENV-4), each exhibiting significant genetic variation. Within these serotypes, multiple genotypes have been identified, often associated with specific geographic regions. Studies have shown that the introduction of new genotypes into a population can lead to increased disease severity and outbreak frequency [5].

Similarly, chikungunya virus (CHIKV) has undergone notable genetic evolution, particularly the emergence of the East/Central/South African (ECSA) genotype, which acquired mutations enhancing its transmission by *Aedes albopictus*. This adaptation has facilitated the spread of chikungunya into previously non-endemic regions, including rural India (Table 3).

Table 3. Genetic characteristics of major arboviruses.

Virus	Genome type	Genome size (kb)	Key proteins	Notable genetic features
Dengue Virus	+ssRNA	~11	E, NS1, NS5	4 serotypes, high mutation rate.
Chikungunya Virus	+ssRNA	~12	E1, E2, nsP1–nsP4	E1 mutation enhances vector spread.
Japanese Encephalitis	+ssRNA	~11	E, NS3, NS5	Genotype shifts linked to outbreaks.
Zika Virus	+ssRNA	~10.7	prM, E, NS1	Neurotropic mutations.
KFD Virus	+ssRNA	~11	E, NS proteins	Tick-borne adaptation.

The genomic evolution of arboviruses is influenced by selective pressures exerted by hosts, vectors, and environmental conditions. In rural India, these pressures are particularly dynamic due to seasonal variations, agricultural practices, and biodiversity. As a result, localized viral strains often exhibit unique genetic signatures, which may not be detected by standard diagnostic assays [6].

CLASSIFICATION AND TYPES OF ARBOVIRUSES: TAXONOMICAL CLASSIFICATION

Arboviruses are taxonomically diverse and belong to multiple viral families based on their genomic structure, replication strategy, and antigenic properties. The classification of arboviruses is primarily based on the guidelines provided by the International Committee on Taxonomy of Viruses, which standardizes viral nomenclature and taxonomy [7].

Broadly, arboviruses affecting humans in India can be classified into the following major families:

- Flaviviridae.
- Togaviridae.
- Bunyaviridae (now reclassified under *Peribunyaviridae* and related families).
- Reoviridae.

Each family comprises viruses with distinct genomic features, transmission patterns, and pathogenic profiles (Table 4).

Table 4. Taxonomical classification of key arboviruses.

Family	Genus	Representative virus	Genome type	Vector type
Flaviviridae	<i>Flavivirus</i>	Dengue virus, Zika virus, Japanese encephalitis virus	+ssRNA	Mosquito (<i>Aedes</i> , <i>Culex</i>).
Togaviridae	<i>Alphavirus</i>	Chikungunya virus	+ssRNA	Mosquito (<i>Aedes</i>).
Peribunyaviridae	<i>Orthobunyavirus</i>	La Crosse virus (related group)	-ssRNA	Mosquito.
Reoviridae	<i>Orbivirus</i>	Bluetongue virus (animal arbovirus)	dsRNA	Midges.

Among these families, *Flaviviridae* is the most clinically significant in India due to the high burden of diseases such as dengue and Japanese encephalitis. These viruses share structural similarities, including an envelope protein that plays a critical role in host cell entry and immune recognition.

CLASSIFICATION AND TYPES OF ARBOVIRUSES (PART II): BASED ON TRANSMISSION CYCLES

Apart from taxonomical classification, arboviruses can also be categorized based on their transmission cycles, which are crucial for understanding their epidemiology and control strategies [8].

Urban Transmission Cycle

In this cycle, the virus is transmitted between humans and vectors, primarily mosquitoes such as *Aedes aegypti*. This cycle is typical for viruses, like dengue and Zika, which thrive in densely populated areas but are increasingly extending into rural settings.

Sylvatic (Jungle) Cycle

In this cycle, the virus circulates between wild animals (e.g., primates, birds) and vectors. Humans become incidental hosts when they enter forested areas. Kyasanur Forest Disease and certain strains of chikungunya follow this pattern [9].

Rural/Agricultural Cycle

This is particularly relevant in India, where viruses like Japanese encephalitis circulate between pigs, birds, and *Culex* mosquitoes in agricultural environments such as rice fields.

Mechanism of Action: Viral Entry and Host Cell Interaction

The pathogenesis of arboviruses begins with their entry into the human host through the bite of an infected arthropod vector, most commonly mosquitoes. Once introduced into the bloodstream, the virus targets specific host cells such as dendritic cells, macrophages, and endothelial cells. The mechanism of viral entry is primarily mediated by interactions between viral surface proteins and host cell receptors [10].

In viruses belonging to the *Flaviviridae* family, such as dengue and Zika, the envelope (E) protein plays a crucial role in host cell attachment and membrane fusion. The virus binds to cellular receptors, like DC-SIGN, heparan sulfate, or TIM receptors, facilitating receptor-mediated endocytosis.

Following endocytosis, the acidic environment within endosomes triggers conformational changes in viral proteins, leading to fusion of the viral envelope with the endosomal membrane. This process releases the viral RNA genome into the cytoplasm, marking the initiation of the replication cycle [11].

Mechanism of Action: Viral Replication and Protein Synthesis

Once inside the host cell, arboviruses utilize the host's cellular machinery for replication and protein synthesis. Most arboviruses possess positive-sense single-stranded RNA (+ssRNA), which can directly function as messenger RNA (mRNA).

The replication process involves the following key steps:

- *Translation of Viral RNA:* The viral genome is translated into a single polyprotein.
- *Proteolytic Cleavage:* Host and viral proteases cleave the polyprotein into functional structural and non-structural proteins.
- *Formation of Replication Complex:* Non-structural proteins form a replication complex on intracellular membranes.
- *RNA Replication:* Synthesis of a complementary negative-strand RNA, which serves as a template for producing new positive-strand genomes [12].

Genomic Characterization Techniques: Sample Collection and Nucleic Acid Extraction

Genomic characterization of arboviruses begins with the collection of appropriate biological samples from infected individuals, vectors, or reservoir hosts. In rural India, commonly collected clinical samples include blood, serum, plasma, cerebrospinal fluid (CSF), and occasionally tissue samples in severe or fatal cases. Additionally, entomological surveillance involves the collection of mosquito vectors such as *Aedes* and *Culex* species [13].

The integrity and quality of the sample are critical for accurate genomic analysis. Improper handling, delays in transport, or exposure to unfavorable temperatures can lead to RNA degradation, significantly affecting sequencing outcomes. Therefore, maintaining a cold chain and using RNA stabilization reagents are essential practices (Table 5).

Table 5. Types of samples used for arbovirus genomic analysis.

Sample type	Source	Advantages	Limitations
Blood/Serum	Human patients	High viral load during acute phase	Limited detection window.
CSF	Neurological cases	Useful for neurotropic viruses	Invasive collection.

Mosquito Pools	Vectors	Early detection of circulation	Requires entomological expertise.
Tissue Samples	Severe cases	Detailed pathology correlation	Rare availability.

Genomic Characterization Techniques (Part II): Reverse Transcription and PCR-Based Methods

Since most arboviruses possess RNA genomes, reverse transcription is a crucial step in converting viral RNA into complementary DNA (cDNA). This process is catalyzed by reverse transcriptase enzymes and forms the basis for several downstream molecular techniques [14].

One of the most widely used methods is Reverse Transcription Polymerase Chain Reaction (RT-PCR), which allows amplification of specific viral gene regions. Real-time RT-PCR (qRT-PCR) further enhances sensitivity and enables quantitative detection of viral load (Table 6).

Table 6. Comparison of PCR-based techniques.

Method	Sensitivity	Specificity	Quantification	Application
Conventional RT-PCR	Moderate	High	No	Detection of known viruses.
qRT-PCR	High	Very High	Yes	Viral load estimation.
Multiplex PCR	High	High	Limited	Detection of multiple viruses.

Genomic Characterization Techniques: Next-Generation Sequencing (NGS) and Whole Genome Sequencing (WGS)

Next-generation sequencing (NGS) has revolutionized the field of viral genomics by enabling high-throughput, rapid, and comprehensive analysis of viral genomes. Unlike PCR-based methods, NGS does not require prior knowledge of the viral sequence, making it particularly useful for detecting novel or emerging arboviruses.

In NGS, fragmented DNA or cDNA libraries are prepared and sequenced in parallel, generating millions of short reads. These reads are then assembled using bioinformatics tools to reconstruct the complete viral genome (Table 7).

Table 7. Advantages of NGS in arbovirus genomics.

Feature	Benefit
High Throughput	Simultaneous sequencing of multiple samples.
Comprehensive Analysis	Detection of known and novel viruses.
Mutation Identification	Detection of single nucleotide variations.
Phylogenetic Insights	Understanding evolutionary relationships.
Co-infection Detection	Identification of multiple pathogens.

RESULTS AND DISCUSSION

Genomic characterization studies of emerging arboviruses in rural India have provided critical insights into viral diversity, mutation patterns, and transmission dynamics. Analysis of sequencing data obtained through RT-PCR, next-generation sequencing (NGS), and whole genome sequencing (WGS) has revealed the presence of multiple co-circulating arboviral strains with significant genetic variability [15].

One of the key findings across multiple studies is the co-circulation of different serotypes and genotypes, particularly in dengue virus. Rural regions, previously considered low-risk zones, are now showing genotypic diversity comparable to urban areas, indicating widespread viral dissemination. Phylogenetic analyses have demonstrated that many rural strains cluster closely with urban and even international isolates, suggesting frequent viral introductions through human movement and vector migration.

Another significant observation is the identification of novel mutations in key viral genes, particularly in envelope and non-structural protein regions. These mutations are associated with:

- Increased viral infectivity.
- Enhanced vector adaptability.
- Potential immune evasion mechanisms.

For example, mutations in the E1 gene of chikungunya virus have been linked to improved transmission by *Aedes albopictus*, a vector increasingly prevalent in rural environments. Similarly, genomic variations in dengue virus NS1 and NS5 proteins may influence replication efficiency and disease severity (Table 8).

Table 8. Key genomic findings in rural arbovirus studies.

Virus	Genomic observation	Implication
Dengue Virus	Multiple serotypes (DENV-1 to DENV-4)	Increased risk of severe infection (ADE).
Chikungunya Virus	E1 mutation (A226V)	Enhanced vector transmission.
Japanese Encephalitis	Genotype shifts (III to I)	Altered epidemiology.
Zika Virus	Asian lineage detection	Emerging threat in rural areas.
KFD Virus	Localized genetic clusters	Region-specific adaptation.

Metagenomic studies have also revealed co-infections and the presence of previously undetected viral agents, emphasizing the complexity of arboviral ecology in rural settings. These findings highlight the limitations of conventional diagnostic approaches and reinforce the need for comprehensive genomic surveillance [16, 17].

Furthermore, genomic data have enabled tracking of outbreak origins and transmission pathways, allowing health authorities to implement targeted interventions. For instance, sequence-based epidemiology has been used to trace dengue outbreaks to specific geographic clusters and identify hotspots of transmission.

Despite these advancements, several challenges persist:

- Limited genomic data from rural and remote areas.
- Inadequate laboratory infrastructure.
- High cost and technical complexity of sequencing technologies.
- Lack of trained personnel in bioinformatics.

These challenges contribute to underrepresentation of rural viral diversity in national and global databases [18].

CONCLUSION

The genomic characterization of emerging arboviruses in rural India has emerged as a vital tool in understanding the evolving landscape of vector-borne diseases. The integration of advanced sequencing technologies with epidemiological surveillance has revealed significant genetic diversity, frequent mutations, and complex transmission dynamics of arboviruses in rural settings.

Key conclusions drawn from this review include:

- Rural India is no longer a peripheral zone but a significant hotspot for arboviral emergence and evolution.
- Genomic studies have identified novel mutations and co-circulating strains, which may influence disease severity and transmission.
- Next-generation sequencing and bioinformatics play a crucial role in early detection, outbreak tracking, and vaccine development.
- There exists a critical need to expand genomic surveillance infrastructure in rural areas.

In conclusion, genomic characterization not only enhances our understanding of arboviral biology but also serves as a cornerstone for developing targeted interventions, improving diagnostic accuracy,

and strengthening outbreak preparedness in rural India. Future research should focus on real-time genomic surveillance, integration with artificial intelligence, and development of cost-effective sequencing technologies to ensure equitable healthcare access across all regions.

REFERENCES

1. Akiner MM, Demirci B, Babuadze G, Robert V, Schaffner F. Spread of the invasive mosquitoes *Aedes aegypti* and *Aedes albopictus* in the Black Sea region increases risk of chikungunya, dengue, and Zika outbreaks in Europe. *PLoS Negl Trop Dis*. 2016;10(4):e0004664.
2. Rogers DJ, Randolph SE. Climate change and vector-borne diseases. *Adv Parasitol*. 2006;62:345–81.
3. Centers for Disease Control and Prevention (CDC). West Nile virus disease and other arboviral diseases—United States, 2010. *MMWR Morb Mortal Wkly Rep*. 2011;60(30).
4. Rai TK, Chakravarty J, Kashyap S, Chatterjee S, Tiwari VD, Rai UG, et al. Etiology of acute encephalitis syndrome in adults in a tertiary care center in Eastern Uttar Pradesh. *Am J Trop Med Hyg*. 2024;112(1):194.
5. Priskilla JJ, Kulandaisamy AM, Appadurai DR, Srirama S, Ananganallur Nagarajan S, Rahi M. Industrial hotspot: Infestation of invasive *Aedes aegypti* and *Aedes albopictus* in Puducherry, India. *Trop Med Int Health*. 2025;30(8):823–30.
6. Matthews RE. Classification and nomenclature of viruses. *Intervirology*. 1982;17(1):1–99.
7. Weaver SC, Reisen WK. Present and future arboviral threats. *Antiviral Res*. 2010;85(2):328–45.
8. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The global distribution and burden of dengue. *Nature*. 2013;496(7446):504–7.
9. Staples JE, Breiman RF, Powers AM. Chikungunya fever: An epidemiological review of a re-emerging infectious disease. *Clin Infect Dis*. 2009;49(6):942–8.
10. Solomon T, Dung NM, Kneen R, Gainsborough M, Vaughn DW, Khanh VT. Japanese encephalitis. *J Neurol Neurosurg Psychiatry*. 2000;68(4):405–15.
11. Landry ML, George KS. Laboratory diagnosis of Zika virus infection. *Arch Pathol Lab Med*. 2017;141(1):60–7.
12. Shah SZ, Jabbar B, Ahmed N, Rehman A, Nasir H, Nadeem S, et al. Epidemiology, pathogenesis, and control of a tick-borne disease-Kyasanur forest disease: Current status and future directions. *Front Cell Infect Microbiol*. 2018;8:149.
13. Greninger AL. A decade of RNA virus metagenomics is (not) enough. *Virus Res*. 2018;244:218–29.
14. Quick J, Loman NJ, Duraffour S, Simpson JT, Severi E, Cowley L, et al. Real-time, portable genome sequencing for Ebola surveillance. *Nature*. 2016;530(7589):228–32.
15. Metsky HC, Matranga CB, Wohl S, Schaffner SF, Freije CA, Winnicki SM, et al. Zika virus evolution and spread in the Americas. *Nature*. 2017;546(7658):411–5.
16. Hesse RR. Dengue virus evolution and virulence models. *Clin Infect Dis*. 2007;44(11):1462–6.
17. Powers AM. Risks to the Americas associated with the continued expansion of chikungunya virus. *J Gen Virol*. 2015;96(1):1–5.
18. Gire SK, Goba A, Andersen KG, Sealfon RS, Park DJ, Kanneh L, et al. Genomic surveillance elucidates Ebola virus origin and transmission during the 2014 outbreak. *Science*. 2014;345(6202):1369–72.