

Zebrafish in Drug Research: Decoding Biological Mechanisms and Finding Novel Therapeutic Targets

Ashish Kushwaha*

Abstract

*The zebrafish, scientifically known as *Danio rerio*, is a powerful model organism in biological research because of its favorable traits, which include rapid development, genetic tractability, and transparent embryos. Zebrafish were first identified for their use in studies of vertebrate development, but they have since spread to a variety of disciplines, such as pharmacology, clinical research as a disease model, and most notably, drug development. Zebrafish have emerged as a cost-effective and practical alternative to traditional mammalian models in numerous biomedical research areas. Their utility spans a wide range of applications such as lead compound screening, identification of molecular targets, morpholino oligonucleotide-based gene function studies, assay development, and the evaluation of drug toxicity. Due to their genetic similarity to humans, transparent embryos, and rapid development, zebrafish serve as an ideal in vivo model for understanding complex biological processes. This review highlights the key characteristics that contribute to the zebrafish's growing significance in research, especially in the field of pharmacology. It emphasizes their vital role in accelerating drug discovery and development, offering a simplified yet reliable system for studying disease mechanisms, therapeutic responses, and adverse drug reactions, making them an invaluable asset in pharmaceutical research.*

KEYWORDS: Toxicity, zebrafish, pharmacology, drug development, embryos, genetic

INTRODUCTION

Originating in South Asia, the zebrafish has become well-known in several biological research domains such as toxicology, neurology, genetics, developmental biology, and pharmacology. Certain characteristics make it an important model organism [1, 2]. Finding chemical entities with potential for medical use is known as drug discovery. The objective is to find new compounds that address unmet medical needs, with an emphasis on serious illnesses for which there are no effective treatments. Current drugs primarily focus on enzymes and target ion channels, nuclear receptors, and G-protein coupled receptors [3].

Due to things, like unknown variables during lead discovery, compounds may encounter difficulties in development even with promising leads. Failures can be caused by problems such as unacceptable toxicity, inadequate biopharmaceutical qualities, and inadequate in vivo efficacy. Complexity of the synthetic process, unclear toxicological results, and low potency can also impede development [2–4].

*Author for Correspondence

Ashish Kushwaha

E-mail: Kushwahaashish577@gmail.com

Assistant Professor, Department of Pharmacy, JBIT College of Pharmacy, Dehradun, Uttarakhand, India.

Received Date: May 10, 2025

Accepted Date: August 25, 2025

Published Date: December 2, 2025

Citation: Ashish Kushwaha. Zebrafish in Drug Research: Decoding Biological Mechanisms and Finding Novel Therapeutic Targets. Research & Reviews: A Journal of Toxicology. 2025; 15(3): 1–8p.

Since the 1980s, the zebrafish (*Danio rerio*), a commonly used freshwater teleost, has been a popular model in studies. The discovery of millions of zebrafish mutant models with conserved disease genes broadened its applications, which were initially concentrated on developmental biology. With the help of their external embryo development, zebrafish's relevance to inherited and acquired human disorders increased the potential for translational research.

The zebrafish's role in translational research has grown since its 2013 genome sequencing revealed

Genetic Tractability

The genome of zebrafish is well-characterized, and researchers can make precise genetic changes using genetic manipulation methods like CRISPR/Cas9. This makes it easier to research pharmacological targets and pathways [10].

Rapid Development

Zebrafish embryos develop rapidly, with their organs and systems forming in just a few days. This fast-paced development enables scientists to assess drug effects and toxicity across multiple stages, offering a comprehensive view of how drugs interact with biological systems [11].

Regenerative Abilities

Zebrafish possess remarkable regenerative abilities, particularly in restoring fins, heart tissue, and the central nervous system. This capacity makes them a valuable model for studying tissue regeneration and repair mechanisms [12].

Developmental Biology

Zebrafish embryos experience fast embryonic development, offering valuable insights into vertebrate growth processes. Their transparency allows clear observation of organ formation, making them an ideal model for exploring developmental biology.

Disease Modelling

Zebrafish are effectively used to model a wide range of human diseases, including neurological conditions, heart disorders, and cancer. Their physiological resemblance to humans, along with ease of genetic modification, enables researchers to investigate disease processes and evaluate potential treatments.

Toxicology and Drug Screening

Zebrafish models serve as a vital tool in toxicology and drug screening research. They play a key role in evaluating the toxicity of both environmental contaminants and pharmaceutical agents, greatly supporting safety assessment efforts.

ZEBRAFISH MODELS IN DISEASE RESEARCH

Zebrafish serve as an excellent model organism for disease and translational research, witnessing a surge in utilization over the last five years. Table 1 shows Different types of zebrafish models and organ used in disease and activity research.

Table 1. Different types of zebrafish models and organs are used in disease and activity research.

S. N.	Activity or Disease Activity	Organ or Models of Zebrafish
1.	Antibacterial Activity	Embryos and Tissue [13].
2.	CNS Effect of Antimicrobial Drugs	Brain [14].
3.	Seizures Induced (Behavioral Characterization)	Adult Zebra fish [15].
4.	Epigenetics in renal disease	Kidney [16].
5.	Neurotoxic Effects and Neurodegeneration (Behavioral Assessment)	Brain [17].
6.	Senescence Independent Anti-Inflammatory activity	Zebra fish Larvae [18].
7.	Inflammatory Bowel Disease	Intestine [19].
8.	Lipid-Lowering drug Induced Myopathies	Muscle [20].
9.	Cancer therapy Induced Cardiovascular Toxicity	Heart [21].
10.	Glioblastoma Cellular and Molecular Mechanisms	Glial Cells [22].
11.	Toxicity	Embryo [23].
12.	Drug-Induced Liver Injury	Liver [24].
13.	Neuropsychopharmacology & CNS Drug Discovery	Brain [25].
14.	Locomotor Activity	Brain [26].
15.	Optical Mapping of Neuronal Activity During Seizures	Brain [27].

16.	Alzheimer's Disease	Brain [28].
17.	Neutrophilic Inflammation	Larvae Neutrophils [29].
18.	Embryo Toxicity	Zebrafish Blood [30].
19.	Autism Spectrum Disorder (ASD)	Brain [31].
20.	Analysis of the Retina	Eye (Retina) [32].
21.	Thyroid Gland Development and Function	Thyroid gland [33].
22.	Dual Oxidase – Anti-Bacterial Properties	Larvae [34].
23.	A Model of Superinfection of Increased Susceptibility to Bacteria Associated with Neutrophil Death.	Virus-Infected Zebrafish Larvae [35].
24.	Antibiotic Chlortetracycline Causes Transgenerational Immunosuppressant Iinf-Kb:	Larvae [36].
25.	Anticarcinogenic And Antioxidant Action of An Edible Aquatic Flora Jussiaea Repens L.	Larvae [37].
26.	Mucosal Inflammation at The Repository Interface	Zebrafish gills [38].
27.	Quantitative analysis of Protein Turnover and Tissue Regeneration	Tissues [39].
28.	Screening Antimycobacterial Compounds	Naturally Infected Zebrafish Larvae Model [40].
29.	Antifungal Activity Of 2'-Hydroxychalcone Loaded in Nanoemulsion Against Paracoccidioides Spp.	Whole Zebrafish [41].
30.	Influenza A And Exacerbate Disease –Duchenne Muscular Dystrophy	Virus Infection Damage Zebrafish Skeletal Muscle [42].
31.	Spermatogonial Stem Cell Niche Spermatogonial Stem Cell Transplantation	Spermatogonial Stem Cells (SSCS) [43].
32.	Estrogen Signaling Influences Nephron Segmentation	Zebrafish Embryonic Kidney [44].
33.	Exocrine Pancreas Development	Pancreas [45].
34.	Ocular Tuberculosis.	Eyes [46].
35.	Bacterial Meningitis in Streptococcus Agalactiae Infection.	Adult Zebrafish [47].
36.	Visual Of Von-Hippel Lindau Disease	Eye [48].
37.	Attention Deficit Hyperactivity Disorder (ADHD).	Zebrafish [49].

Nutritional Abnormalities in Zebrafish

Zebrafish, whose major organs closely resemble those involved in human metabolic regulation, serve as a valuable model for researching obesity and metabolic disorders. By experimentally altering their blood glucose levels to mimic diabetes, researchers have gained important insights into complications like retinal inflammation. Additionally, studies on osmoregulation have highlighted links to glucose absorption, providing a distinctive approach to exploring diabetic retinopathy.

Frameworks of Inflammatory Pathology in Zebrafish

Zebrafish models are instrumental in exploring inflammation and tissue regeneration, particularly through tail injury-induced acute inflammation. They provide a platform to investigate glucocorticoid activity, anti-inflammatory effects, and the discovery of novel therapeutic agents. Pro-resolution pathways, such as those involving lipoxins, play a key role in resolving inflammation. Moreover, the presence of mast cells in zebrafish, which are comparable to those in mammals, offers valuable insights into inflammatory processes.

Cancer Research in Zebrafish

Zebrafish are highly useful in cancer research, exhibiting spontaneous tumor formation and supporting techniques like genetic engineering, transgenic modelling, allografts, xenografts, and high-throughput genetic screening. Their transparent embryos enable real-time observation of tumor development, offering a distinct advantage for studying cancer biology. These models effectively mimic a range of human cancers and are instrumental in identifying and testing potential therapeutic agents.

CARDIOVASCULAR STUDY USING ZEBRAFISH AS A GENETIC MODEL

Zebrafish present key advantages for investigating cardiovascular genetics, with their transparent embryos allowing for detailed phenotypic screening. The clearly visible embryonic heart facilitates

direct observation of heart development. Genetic screens help identify heart-related abnormalities, enhancing our understanding of circulatory disorders. These models also replicate human responses to cardiac drugs, making them valuable tools for drug development. Additionally, high-throughput assays support rapid and efficient testing of cardiovascular impacts.

Zebrafish in Toxicology and Safety Assessment

Zebrafish are a valuable model for evaluating developmental and reproductive toxicity during drug development, enabling early detection of potential risks. Assessing organ-specific toxicity, especially in the heart and liver, is a key aspect of preclinical testing. As vertebrate organisms, zebrafish play a significant role in *in vivo* toxicity screening, as well as in evaluating drug efficacy and overall safety. Safety pharmacology focuses on identifying adverse effects on the respiratory, neurological, and cardiovascular systems, which can vary depending on the compound's class and mechanism of action.

Embryotoxicity and Teratogenicity

Zebrafish models are instrumental in investigating embryotoxicity and teratogenicity, offering insights into compound-induced structural abnormalities and disease mechanisms. Their fully sequenced genome supports the study of signaling pathways associated with teratogenic effects. The impact of drugs on development and reproduction is thoroughly assessed, following protocols refined since the discovery of thalidomide's harmful effects. The rapid development of zebrafish embryos enables efficient evaluation of viability, morphological defects, and emerging endpoints, moving beyond basic survival analysis.

Cardiotoxicity and Hepatotoxicity

Zebrafish embryos are highly effective in toxicological research, particularly for examining the effects of environmental pollutants and developmental toxins on the cardiovascular system. Investigations involving cigarette smoke and chronic alcohol exposure underscore their value in assessing cardiovascular toxicity. Zebrafish models also play a significant role in evaluating the cardiotoxic effects of human medications, reinforcing their importance in drug safety studies. In hepatotoxicity research, both *in vitro* and *in vivo* approaches are employed, with zebrafish embryos exhibiting gene expression changes comparable to those seen with human liver toxins. The distinctive tubular structure of the zebrafish liver allows for real-time observation of specific cellular responses, aiding in the study of liver damage caused by drugs and environmental agents.

CHALLENGES AND LIMITATIONS

Zebrafish model presents several challenges. While it allows to produce large numbers of embryos at a low cost, issues persist, such as the difficulty in correlating toxic water dosing levels with mammalian plasma concentrations, along with the need for more research on absorption, distribution, metabolism, and excretion (ADME) data. Other limitations include partial genome duplication, which leads to gene redundancy, and a relatively moderate generation time. While zebrafish are convenient for laboratory studies, these challenges, particularly in toxicological research, raise concerns about their applicability to human models.

Future Direction

As a preclinical model, the zebrafish requires further research to pinpoint the molecular targets of tested chemicals, particularly those affecting cardiac-related genes and proteins. A thorough analysis of how these chemicals influence relevant pathways and disrupt associated traits is essential. Investigations should prioritize established molecular targets and signaling pathways, while also remaining open to discovering new ones if needed. Addressing these factors is key to advancing zebrafish research and improving its utility in drug safety evaluation [49].

CONCLUSIONS

The zebrafish, known for its genetic strengths, swift developmental cycle, and impressive regenerative abilities, serves as a vital model in a wide range of biological research fields. It is integral to exploring basic biological functions, modelling diseases, and assessing drug efficacy. Despite its indispensable role, there is

a continuing need to advance welfare practices, refine disease models, and enhance drug screening methods. Sustained efforts in these areas will further the study of *Danio rerio*, offering deeper insights into its biomedical research potential.

Acknowledgment

The authors gratefully acknowledge the departmental authority and the CV Raman Research Wing of JBIT College of Pharmacy for their invaluable support in conducting this study. They also extend their sincere thanks to the anonymous referee for their constructive feedback and insightful suggestions for revision.

REFERENCES

1. Barros TP, Alderton WK, Reynolds HM, Roach AG, Berghmans S. Zebrafish: An emerging technology for in vivo pharmacological assessment to identify potential safety liabilities in early drug discovery. *Br J Pharmacol*. 2008 Aug;154(7):1400–13.
2. MacRae CA, Peterson RT. Zebrafish as a mainstream model for in vivo systems pharmacology and toxicology. *Annu Rev Pharmacol Toxicol*. 2023 Jan 20;63:43–64.
3. Zon LI, Peterson RT. In vivo drug discovery in the zebrafish. *Nat Rev Drug Discov*. 2005 Jan 1;4(1):35–44.
4. Singleman C, Holtzman NG. Growth and maturation in the zebrafish, *Danio rerio*: A staging tool for teaching and research. *Zebrafish*. 2014;11(4):396–406. doi:10.1089/zeb.2014.0976.
5. Strähle U, Grabher C. The zebrafish embryo as a model for assessing off-target drug effects. *Dis Model Mech*. 2010 Oct 28;3(11–12):689–92.
6. MacRae CA, Peterson RT. Zebrafish as tools for drug discovery. *Nat Rev Drug Discov*. 2015 Oct;14(10):721–31.
7. Zon LI, Peterson RT. In vivo drug discovery in the zebrafish. *Nat Rev Drug Discov*. 2005 Jan 1;4(1):35–44.
8. Zanandrea R, Bonan CD, Campos MM. Zebrafish as a model for inflammation and drug discovery. *Drug Discov Today*. 2020 Dec 1;25(12):2201–11.
9. Rubinstein AL. Zebrafish: From disease modeling to drug discovery. *Curr Opin Drug Discov Devel*. 2003 Mar 1;6(2):218–23.
10. Wang X, Zhang JB, He KJ, Wang F, Liu CF. Advances of zebrafish in neurodegenerative disease: From models to drug discovery. *Front Pharmacol*. 2021 Jul 14;12:713963.
11. Khan KM, Collier AD, Meshalkina DA, Kysil EV, Khatsko SL, et al. Zebrafish models in neuropsychopharmacology and CNS drug discovery. *Br J Pharmacol*. 2017 Jul;174(13):1925–44.
12. Xie Y, Meijer AH, Schaaf MJ. Modeling inflammation in zebrafish for the development of anti-inflammatory drugs. *Front Cell Dev Biol*. 2021 Jan 15;8:620984.
13. González-Rentería M, del Carmen Monroy-Dosta M, Guzmán-García X, Hernández-Calderas I. Antibacterial activity of *Lemna minor* extracts against *Pseudomonas fluorescens* and safety evaluation in a zebrafish model. *Saudi J Biol Sci*. 2020 Dec 1;27(12):3465–73.
14. Kotova MM, Galstyan DS, Kolesnikova TO, De Abreu MS, Amstislavskaya TG, Strekalova T, et al. Understanding CNS effects of antimicrobial drugs using zebrafish models. *Vet Sci*. 2023 Jan 29;10(2):96.
15. Mussulini BH, Leite CE, Zenki KC, Moro L, Baggio S, Rico EP, et al. Seizures induced by pentylenetetrazole in the adult zebrafish: A detailed behavioral characterization. *PLoS One*. 2013 Jan 21;8(1):e54515.
16. Söpel N, Müller-Deile J. The zebrafish model to understand epigenetics in renal diseases. *Int J Mol Sci*. 2021 Aug 25;22(17):9152.
17. Lee J, Freeman JL. Zebrafish as a model for investigating developmental lead (Pb) neurotoxicity as a risk factor in adult neurodegenerative disease: A mini-review. *Neurotoxicology*. 2014 Jul 1;43:57–64.
18. Hernández-Silva D, Cantón-Sandoval J, Martínez-Navarro FJ, Pérez-Sánchez H, de Oliveira S, Mulero V, et al. Senescence-independent anti-inflammatory activity of the senolytic drugs dasatinib, navitoclax, and venetoclax in zebrafish models of chronic inflammation. *Int J Mol Sci*. 2022 Sep 9;23(18):10468.
19. Fleming A, Jankowski J, Goldsmith P. In vivo analysis of gut function and disease changes in a zebrafish larvae model of inflammatory bowel disease: A feasibility study. *Inflamm Bowel Dis*. 2010 Jul 1;16(7):1162–72.
20. Asnani A, Moslehi JJ, Adhikari BB, Baik AH, Beyer AM, de Boer RA et al. Preclinical models of cancer therapy-associated cardiovascular toxicity: A scientific statement from the American Heart Association. *Circ Res*. 2021 Jun 25;129(1):e21–34.

21. Reimunde P, Pensado-López A, Carreira Crende M, Lombao Iglesias V, Sánchez L, Torrecilla Parra M, et al. Cellular and molecular mechanisms underlying glioblastoma and zebrafish models for the discovery of new treatments. *Cancers (Basel)*. 2021 Mar 3;13(5):1087. doi:10.3390/cancers13051087. PMID:33803314; PMCID:PMC7956469.
22. He JH, Gao JM, Huang CJ, Li CQ. Zebrafish models for assessing developmental and reproductive toxicity. *Neurotoxicol Teratol*. 2014 Mar 1;42:35–42.
23. Vliegthart AB, Tucker CS, Del Pozo J, Dear JW. Zebrafish as model organisms for studying drug-induced liver injury. *Br J Clin Pharmacol*. 2014 Dec;78(6):1217–27.
24. Khan KM, Collier AD, Meshalkina DA, Kysil EV, Khatsko SL, Kolesnikova T, et al. Zebrafish models in neuropsychopharmacology and CNS drug discovery. *Br J Pharmacol*. 2017 Jul;174(13):1925–44.
25. Selderslaghs IW, Hooyberghs J, De Coen W, Witters HE. Locomotor activity in zebrafish embryos: A new method to assess developmental neurotoxicity. *Neurotoxicol Teratol*. 2010 Jul 1;32(4):460–71.
26. Turrini L, Fornetto C, Marchetto G, Müllenbroich MC, Tiso N, Vettori A, et al. Optical mapping of neuronal activity during seizures in zebrafish. *Sci Rep*. 2017 Jun 8;7(1):3025.
27. Newman M, Ebrahimie E, Lardelli M. Using the zebrafish model for Alzheimer’s disease research. *Front Genet*. 2014 Jun 30;5:189.
28. Renshaw SA, Loynes CA, Trushell DM, Elworthy S, Ingham PW, Whyte MK. A transgenic zebrafish model of neutrophilic inflammation. *Blood*. 2006 Dec 15;108(13):3976–8.
29. Tshering G, Plengsuriyakarn T, Na-Bangchang K, Pimtong W. Embryotoxicity evaluation of atracylodine and β -eudesmol using the zebrafish model. *Comp Biochem Physiol C Toxicol Pharmacol*. 2021 Jan 1;239:108869.
30. Stewart AM, Nguyen M, Wong K, Poudel MK, Kalueff AV. Developing zebrafish models of autism spectrum disorder (ASD). *Prog Neuropsychopharmacol Biol Psychiatry*. 2014 Apr 3;50:27–36.
31. Avanesov A, Malicki J. Analysis of the retina in the zebrafish model. *Methods Cell Biol*. 2010 Jan 1;100:153–204.
32. Porazzi P, Calebiro D, Benato F, Tiso N, Persani L. Thyroid gland development and function in the zebrafish model. *Mol Cell Endocrinol*. 2009 Nov 27;312(1–2):14–23.
33. Flores MV, Crawford KC, Pullin LM, Hall CJ, Crosier KE, Crosier PS. Dual oxidase in the intestinal epithelium of zebrafish larvae has antibacterial properties. *Biochem Biophys Res Commun*. 2010 Sep 10;400(1):164–8.
34. Boucontet L, Passoni G, Thiry V, Maggi L, Herbomel P, Levraud JP, et al. A model of superinfection of virus-infected zebrafish larvae: Increased susceptibility to bacteria associated with neutrophil death. *Front Immunol*. 2018 May 24;9:1084.
35. Qiu W, Chen B, Tang L, Zheng C, Xu B, Liu Z, et al. Antibiotic chlortetracycline causes transgenerational immunosuppression via NF- κ B. *Environ Sci Technol*. 2022 Mar 14;56(7):4251–61.
36. Rajiv C, Roy SS, Tamreihao K, Kshetri P, Singh TS, Sanjita Devi H, et al. Anticarcinogenic and antioxidant action of an edible aquatic flora *jussiaea repens* L. using in vitro bioassays and in vivo zebrafish model. *Molecules*. 2021 Apr 15;26(8):2291.
37. Progetzky F, Cook HT, Lamb JR, Bugeon L, Dallman MJ. Mucosal inflammation at the respiratory interface: A zebrafish model. *Am J Physiol Lung Cell Mol Physiol*. 2016 Mar 15;310(6):L551–61.
38. Westman-Brinkmalm A, Abramsson A, Pannee J, Gang C, Gustavsson MK, von Otter M, et al. SILAC zebrafish for quantitative analysis of protein turnover and tissue regeneration. *J Proteomics*. 2011 Dec 21;75(2):425–34.
39. Dalton JP, Uy B, Okuda KS, Hall CJ, Denny WA, Crosier PS, et al. Screening of antimycobacterial compounds in a naturally infected zebrafish larvae model. *J Antimicrob Chemother*. 2016 Oct 22;dkw421. PMID:30779935.
40. Medina-Alarcon KP, L Singulani JD, Dutra LA, S Pitanguí ND, Pereira-da-Silva MA, Dos Santos MB, et al. Antifungal activity of 2'-hydroxychalcone loaded in nanoemulsion against *Paracoccidioides* spp. *Future Microbiol*. 2020 Jan;15(1):21–33.
41. Goody M, Jurczynszak D, Kim C, Henry C. Influenza A virus infection damages zebrafish skeletal muscle and exacerbates disease in zebrafish modeling Duchenne muscular dystrophy. *PLoS Curr*. 2017 Oct 25;9.

42. Nobrega RH, Greebe CD, van de Kant H, Bogerd J, de Franca LR, Schulz RW. Spermatogonial stem cell niche and spermatogonial stem cell transplantation in zebrafish. *PLoS One*. 2010 Sep 20;5(9):e12808.
43. Wesselman HM, Gatz AE, Pfaff MR, Arceri L, Wingert RA. Estrogen signaling influences nephron segmentation of the zebrafish embryonic kidney. *Cells*. 2023 Feb 20;12(4):666.
44. Yee NS, Lorent K, Pack M. Exocrine pancreas development in zebrafish. *Dev Biol*. 2005 Aug 1;284(1):84–101.
45. Damera SK, Panigrahi RK, Mitra S, Basu S. Role of extracellular Mycobacteria in blood-retinal barrier invasion in a zebrafish model of ocular TB. *Pathogens*. 2021 Mar 13;10(3):333.
46. Patterson H, Saralahti A, Parikka M, Dramsi S, Trieu-Cuot P, Poyart C, et al. Adult zebrafish model of bacterial meningitis in *Streptococcus agalactiae* infection. *Dev Comp Immunol*. 2012 Nov 1;38(3):447–55.
47. Ward R, Ali Z, Slater K, Reynolds AL, Jensen LD, Kennedy BN. Pharmacological restoration of visual function in a zebrafish model of von-Hippel Lindau disease. *Dev Biol*. 2020 Jan 15;457(2):226–34.
48. Fontana BD, Franscescon F, Rosenberg DB, Norton WHJ, Kalueff AV, Parker MO. Zebrafish models for attention deficit hyperactivity disorder (ADHD). *Neurosci Biobehav Rev*. 2019 May;100:9–18. doi:10.1016/j.neubiorev.2019.02.009. Epub 2019.
49. Collier AD, Abdulai AR, Leibowitz SF. Utility of the zebrafish model for studying neuronal and behavioral disturbances induced by embryonic exposure to alcohol, nicotine, and cannabis. *Cells*. 2023 Oct 23;12(20):2505. doi:10.3390/cells12202505.